AmpliSens® *HCV*-Monitor-FRT PCR kit

RUO

For Professional Use Only

Instruction Manual

KEY TO SYMBOLS USED

REF	Catalogue number	⚠	Caution
LOT	Batch code	Σ	Contains sufficient for <n> tests
RUO	Research use only		Use-by Date
VER	Version		Consult instructions for use
	Temperature limit		Keep away from sunlight
	Manufacturer	NCA	Negative control of amplification
	Date of manufacture	C–	Negative control of extraction
PCE	Positive Control of Extraction	C+	Positive control of amplification
		IC	Internal control

1. INTENDED USE

AmpliSens® *HCV*-Monitor-FRT PCR kit is an *in vitro* nucleic acid amplification test for quantitative detection of *hepatitis C virus* (*HCV*) RNA in biological material (blood plasma) using real-time hybridization-fluorescence detection of amplified products.

NOTE: For research use only. Not for diagnostic procedures.

2. PRINCIPLE OF PCR DETECTION

Principle of testing is based on the RNA extraction from the samples of test material with the exogenous internal control sample (Internal Control *ICZ*-rec (IC)), RNA reverse transcription and simultaneous amplification of cDNA fragments with hybridization-fluorescence detection. Exogenous internal control (Internal Control *ICZ*-rec (IC)) allows to control all PCR-analysis stages of each individual sample and to identify possible reaction inhibition.

AmpliSens® *HCV*-Monitor-FRT PCR kit uses "hot-start", which greatly reduces frequency of nonspecifically primed reactions. "Hot-start" is guaranteed by using chemically modified polymerase (TaqF). Chemically modified polymerase (TaqF) is activated by heating at 95 °C for 15 min.

The results of amplification are registered in the following fluorescence channels:

Table 1

Channel for fluorophore	FAM	JOE
cDNA-target	IC cDNA	*HCV* cDNA
Target gene	Artificially synthesized sequence	5'UTR

3. CONTENT

AmpliSens® *HCV*-Monitor-FRT PCR kit is produced in 2 forms:

Form 1: PCR kit variant FRT, Negative Control (C–) (4 tubes), ***HCV-Q* calibration kit**, REF R-V1-MC(RG,iQ,Mx,Dt)-CE,

Form 2: PCR kit variant FRT, Negative Control (C–) (4 tubes), ***HCV-Q* calibration kit in bulk**[1], REF R-V1-MC(RG,iQ,Mx,Dt)-CE-B.

PCR kit variant FRT includes:

Reagent		Description	Volume, ml	Quantity
DTT frozen-dried		white powder	–	4 tubes
RT-PCR-mix-1-FL *HCV*		clear liquid from colorless to light lilac colour	0.3	4 tubes
RT-PCR-mix-2-FEP/FRT		colorless clear liquid	0.2	4 tubes
Polymerase (TaqF)		colorless clear liquid	0.02	4 tubes
TM-Revertase (MMlv)		colorless clear liquid	0.01	4 tubes
DNA calibrators	**PIC1 *HCV***	colorless clear liquid	0.1	4 tubes
	PIC2 *HCV*	colorless clear liquid	0.1	4 tubes
Buffer for elution		colorless clear liquid	1.2	4 tubes
Positive Control-1-*HCV**		colorless clear liquid	0.06	4 tubes
Positive Control-2-*HCV**		colorless clear liquid	0.06	4 tubes
Internal Control *ICZ*-rec (IC)*		colorless clear liquid	0.28	4 tubes

***** must be used in the extraction procedure (see section 8.1 for details).

PCR kit variant FRT is intended for 80 reactions, including controls and calibrators.

The following reagent is additionally included in the PCR kit variant FRT:

Reagent	Description	Volume, ml	Quantity
Negative Control (C-)*	colorless clear liquid	1.2	4 tubes

***** must be used in the extraction procedure as Negative Control of Extraction.

***HCV-Q* calibration kit** includes:

Reagent	Description	Volume, ml	Quantity
Calibrator *HCV*-Q	yellow powder	–	1 tube
Solvent Q	colorless clear liquid	1.2	3 tubes

PCR kit also includes:
The software in Microsoft® Excel format for processing of data and generation of results.

4. ADDITIONAL REQUIREMENTS

- DNA/RNA extraction kit.
- Sterile RNase-free/DNase-free pipette tips with filters up to 200 µl and up to 1000 µl.
- Tube racks.
- Vortex mixer.
- Desktop microcentrifuge up to 12,000 g (suitable for Eppendorf tubes).
- PCR box.
- Real-time instruments (for example, Rotor-Gene 3000/6000 (Corbett Research, Australia); iCycler iQ5 (Bio-Rad, USA); Mx3000P (Stratagene, USA).
- Disposable polypropylene tubes:
  a) thin-walled 0.2-ml PCR tubes with optical transparent domed or flat caps or strips of eight 0.2-ml tubes with optical transparent caps if a plate-type instrument is used;
  b) thin-walled 0.2-ml PCR tubes with flat caps or strips of four 0.1-ml Rotor-Gene PCR tubes if a rotor-type instrument is used.
- Pipettes (adjustable).
- Refrigerator for 2-8 °C.
- Deep-freezer at the temperature from minus 24 to minus 16 °C.
- Reservoir for used tips.
- Disposable powder-free gloves and a laboratory coat.

5. GENERAL PRECAUTIONS

The user should always pay attention to the following:

- Use sterile pipette tips with aerosol filters and use a new tip for every procedure.
- Store all extracted positive material (samples, controls and amplicons) away from all other reagents and add it to the reaction mix in a distantly separated facility.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Use disposable protective gloves and laboratory cloths, and protect eyes while samples and reagents handling. Thoroughly wash hands afterwards.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use the PCR kit if the internal packaging was damaged or its appearance was changed.
- Do not use the PCR kit if the transportation and storage conditions according to the Instruction Manual were not observed.
- Do not use a kit after its expiration date.
- Dispose of all specimens and unused reagents in accordance with local regulations.
- Samples should be considered potentially infectious and handled in biological cabinet in compliance with appropriate biosafety practices.
- Clean and disinfect all sample or reagent spills using a disinfectant, such as 0.5% sodium hypochlorite or another suitable disinfectant.
- Avoid inhalation of vapors, samples and reagents contact with the skin, eyes, and mucous membranes. Harmful if swallowed. If these solutions come into contact, rinse the injured area immediately with water and seek medical advice if necessary.
- While observing the conditions of transportation, operation and storage, there are no risks of explosion and ignition.
- Safety Data Sheets (SDS) are available on request.
- The PCR kit is intended for single use for PCR analysis of specified number of samples (see the section "Content").
- The PCR kit is ready for use in accordance with the Instruction Manual. Use the PCR kit strictly for intended purpose.
- Use of this product should be limited to personnel trained in DNA amplification techniques.
- Workflow in the laboratory must be one-directional, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment and reagents in the area where the previous step was performed.

⚠ Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

[1] In bulk form contains unlabeled tubes. Tubes with identical reagent are packed in one bag with label.

6. SAMPLING AND HANDLING

NOTE: Obtaining biological material samples for PCR-analysis, transportation, and storage are described in manufacturer's handbook [1]. It is recommended that this handbook is read before starting work.

AmpliSens® *HCV*-Monitor-FRT PCR kit is intended for the analysis of RNA extracted with RNA extraction kits from the biological material (blood plasma):

Blood samples are collected in the morning on an empty stomach into the tube with EDTA solution as the anticoagulant. Several times invert the closed tubes to ensure proper mixing. To collect plasma, centrifuge the tubes with blood at 800–1600 g for 20 min within 6 h after blood taking. Remove obtained plasma and transfer to the new tubes.

The blood serum may also be used in some cases. The analytical sensitivity of the reagent kit is retained for this material; however, the clinical sensitivity may be significantly decreased as a result of viral particles precipitation during blood clot retraction.

NOTE: The peripheral blood plasma samples and the blood serum samples can be stored for no longer than 3 days at a temperature range from 2 to 8 °C or, if stored at the temperature not more than minus 68 °C, for a long time.

7. WORKING CONDITIONS

AmpliSens® *HCV*-Monitor-FRT PCR kit should be used at the temperature from 20 to 28 °C and relative humidity from 15 to 75 %.

8. PROTOCOL

8.1 RNA extraction

It is recommended to use the following nucleic acid extraction kits:

- **RIBO-prep,** REF K2-9-Et-100-CE,
- **MAGNO-sorb**, REF K2-16-1000-CE.
- NucliSENS easyMAG, automated nucleic acid extraction system (bioMérieux, France) (See Guidelines [2] for details).

NOTE: **If using RIBO-prep kit** extract the RNA/DNA according to the manufacturer's protocol taking into account following additions and improvements:

- If a large number of samples is being tested, it is acceptable to mix the **Solution for Lysis** and the **Internal Control** in a separate sterile flask (based on addition of **300 µl** of **Solution for Lysis** and **10 µl** of **Internal Control** per one sample), followed by a transfer of **300 µl** of the prepared mix into each of the previously prepared **1.5 ml** tubes
- For each panel it is necessary to set up **two** positive controls of extraction – PCE-1 and PCE-2. To the tube labelled PCE-1 add **90 µl** of **Negative Control** and **10 µl** of **Positive Control-1-*HCV***, to the tube labelled PCE-2 add **90 µl** of **Negative Control** and **10 µl** of **Positive Control-2-*HCV***. Close the lids and vortex the tubes.
- Centrifuge at 12,000 g throughout the extraction procedure

NOTE: **If using the MAGNO-sorb kit** extract RNA/DNA according to the manufacturer's protocol taking into account following additions and improvements:

- In case of RNA extraction from blood plasma sample of 1000 µl, the volume of the **Internal Control *ICZ*-rec** required for 24-tube panel is **0.28 ml**. In case of other panels and RNA extraction from blood plasma sample of 200 µl see the MAGNO-sorb instruction manual.
- To prepare the Positive Control of Extraction 1 (**PCE-1**), add **90 µl** of the **Negative Control (C–)** sample and **10 µl** of the **Positive Control-1-*HCV*** sample to the new tube containing **Lysis Solution MAGNO-sorb**.
- To prepare the Positive Control of Extraction 2 (**PCE-2**), add **90 µl** of the **Negative Control (C–)** sample and **10 µl** of the **Positive Control-2-*HCV*** sample to the new tube containing **Lysis Solution MAGNO-sorb**.
- To prepare the Negative Control of Extraction (**C–**), add **100 µl** of the **Negative Control (C–)** sample to the new tube containing **Lysis Solution MAGNO-sorb**.
- The volume of **Buffer for elution** required for extraction from both 1000 and 200 µl of blood plasma samples is **70 µl**.

8.1.1 Calibration and calculation of the coefficient B using *HCV-Q* calibration kit

NOTE: If coefficient B for the extraction kit/automatic platform is not specified in the *Important Product Information Bulletin* enclosed in the PCR kit, the calibration for calculation of coefficient B should be carried out by oneself with the aid of the ***HCV-Q*** calibration kit included in this PCR kit. See below for details.

The calibration procedure is necessary to define Coefficient B and it is performed during the first PCR run for the given lot. Calibration is performed only once for each new lot of the **AmpliSens® *HCV*-Monitor-FRT** PCR kit and is conducted using the same RNA extraction kit/automatic station as was used in the PCR assay. To carry out calibration, it is necessary to analyse 5 extra samples: the repeat of Positive Control-1-*HCV*, the repeat of Positive Control-2-*HCV*, and calibrator *HCV*-Q in triplicate.

Calibrator *HCV*-Q preparation

1. Vortex the tube with **calibrator *HCV*-Q**, gently open the tube, and add **400 µl of solvent Q** avoiding the contents spraying. Use tips with filters.
2. Close the tube and incubate it at room temperature for 20 min vortexing periodically.
3. Once the contents are fully dissolved, vortex the tube for 3-5 s to make sure that there are no drops on the caps of the tube.

Perform calibration with the same RNA extraction kit used in the PCR assay.

NOTE: Extract the RNA according to the manufacturer's protocol.

Transfer 10 µl of **Internal Control *ICZ*-rec (IC)** (per one sample) to samples or to **Lysis solution** before extraction.

In case of extracting from 100 µl of plasma, add dissolved **calibrator *HCV*-Q** to three tubes for RNA extraction (100 µl per each tube).

In case of extracting from any other plasma volume (100 – 1000 µl), transfer dissolved **calibrator *HCV*-Q** to three tubes for RNA extraction (100 µl per each tube) and add **solvent Q** up to the extraction volume (for example, if the extraction volume is 1 ml then add 100 µl of **calibrator *HCV*-Q** and 900 µl of solvent Q).

When extraction is completed, perform PCR as described in this instruction manual.

Use the mean concentration values obtained in the channels for the FAM and JOE fluorophores for three repeats with **calibrator *HCV*-Q** for calculation of coefficient B using the following formula:

$$\textbf{Coefficient B} = \frac{\text{IC cDNA copies in calibrator } HCV\text{-Q (FAM channel)}}{HCV \text{ cDNA copies in calibrator } HCV\text{-Q (JOE/HEX channel)}} \times \text{coefficient C}$$

Coefficient C is specified in the *Important Product Information Bulletin* enclosed in the **AmpliSens® *HCV*-Monitor-FRT** PCR kit.

NOTE: The calculated value of coefficient B should be within range specified in the *Important Product Information Bulletin* enclosed to the applied PCR kit lot.

Write down the coefficient B value in *the Important Product Information Bulletin* enclosed with the given lot of the PCR kit and use it for concentrations calculation of biological and control samples (See the Data Analysis section).

Also see the Guidelines [2] to **AmpliSens® *HCV*-Monitor-FRT** PCR kit.

Write down the calculated values for Positive Control-1-*HCV* and Positive Control-2-*HCV* in the *Important Product Information Bulletin* enclosed with the given lot of the PCR kit.

Determine the mean value for both Positive Contorl-1-*HCV* and for Positive Control-2-*HCV*.

Set the acceptable value range for both Positive Contorl-1-*HCV* and for Positive Control-2-*HCV* as follows: from *"calculated mean value"/3* to *"calculated mean value" x 3.*

For example,

the calculated values for Positive Contorl-1-*HCV* in two replicates are 500,000 IU/ml and 700,000 IU/ml;

the calculated mean value for Positive Contorl-1-*HCV* is 600,000 IU/ml;

the acceptable value range for Positive Contorl-1-*HCV* varies from 200,000 to 1,800,000 IU/ml.

Write down the calculated acceptable value range for Positive Control-1-*HCV* and for Positive Control-2-*HCV* in the *Important Product Information Bulletin,* and use it to verify further assays conducted using this lot of the PCR kit. (See 9. Data Analysis section)

8.2 Preparing reverse transcription and PCR

The total reaction volume is **50 µl**, the volume of RNA sample is **25 µl**.

8.2.1 Preparing tubes for RT-PCR

NOTE: All components of the reaction mix should be mixed just before use. See Table 2 for the reaction mixture preparation scheme.

1. Before starting work, thaw and thoroughly vortex all reagents of the kit. Make sure that there are no drops on the caps of the tubes.
2. Take the required number of the PCR tubes for amplification for the biological and control samples (including 3 controls of extraction and 4 calibrators). The type of tubes depends on the PCR instrument used for analysis.
3. To prepare the **reaction mixture**:
   – add the entire contents of the tube with **RT-PCR-mix-2-FEP/FRT** to the tube with **DTT dried-frozen**. Thoroughly vortex and make sure there are no drops on the caps of the tube. Store the prepared mixture at 2–8 °C for no longer than 2 weeks.
   – mix reagents **15 µl of RT-PCR-mix-1-FL *HCV***, **10 µl of the mixture of RT-PCR-mix-2-FEP/FRT and DTT frozen-dried, 1.0 µl of polymerase (TaqF) and 0.5 µl of TM-Revertase (MMlv)** per one reaction in a new sterile tube. Thoroughly vortex the mixture, make sure that there are no drops on the caps of the tubes.

It is recommended that the reaction mixture for 20 reactions is prepared in case of extraction from 12 to 16 samples (two NucliSENS easyMAG arrays). To do this, into the tube with **DTT frozen-dried** transfer the entire contents of the tubes with **RT-PCR-mix-2-FEP/FRT**, **RT-PCR-mix-1-FL *HCV*, polymerase (TaqF), and TM-Revertase (MMlv)**. Do not store the prepared mixture!

Table 2

Scheme of reaction mixture preparation

			Reagent volumes per number of samples specified plus 1 extra reaction			
Reagent volume per one reaction, µl			**15.00**	**10.00**	**1.00**	**0.5**
Number of biological samples	Number of extracted sampled[2]	Number of samples analyzed in PCR[3]	RT-PCR-mix-1-FL *HCV*	Mixture of RT-PCR-mix-2-FEP/FRT and DTT frozen dried	Polymerase (TaqF)	TM-revertase (MMlv)
3	**6**	**10**	**165**	**110**	**11**	**5.5**
4	**7**	**11**	**180**	**120**	**12**	**6**
5	**8[4]**	**12**	**195**	**130**	**13**	**6.5**
6	**9**	**13**	**210**	**140**	**14**	**7**
7	**10**	**14**	**225**	**150**	**15**	**7.5**
8	**11**	**15**	**240**	**160**	**16**	**8**
9	**12[5]**	**16**	**255**	**170**	**17**	**8.5**
10	**13**	**17**	**270**	**180**	**18**	**9**
11	**14**	**18**	**285**	**190**	**19**	**9.5**
12	**15**	**19**	**Entire contents of the tube**	**Entire contents of the tube**	**Entire contents of the tube**	**Entire contents of the tube**
13	**16[6]**	**20**	**Entire contents of the tube**	**Entire contents of the tube**	**Entire contents of the tube**	**Entire contents of the tube**

4. Transfer **25 µl** of the prepared reaction mixture to each PCR tube. Discard the unused reaction mixture.
5. Using tips with filters add **25 µl** of biological **RNA samples**. Thoroughly mix by pipetting. Avoid formation of air bubbles.

NOTE: Avoid transferring of the sorbent together with the RNA sample in case of extraction with NucliSENS easyMAG automated nucleic acid extraction system or MAGNO-sorb kit.

6. Carry out the control amplification reactions:

PCE-1 – Add **25 µl of RNA sample** extracted from Positive Control-1-*HCV* to the tube labelled **PCE-1** (Positive Control of Extraction).

PCE-2 – Add **25 µl of RNA sample** extracted from Positive Control-2-*HCV* to the tube labelled **PCE-2** (Positive Control of Extraction).

C– – Add **25 µl of RNA sample** extracted from Negative Control (C–) to the tube labelled **C–** (Negative control of Extraction).

$C+_1$ – Add **DNA calibrator PIC1 *HCV*** to the two tubes labelled $C+_1$ (**25 µl per each tube)**.

$C+_2$ – Add **DNA calibrator PIC2 *HCV*** to the two tubes labelled $C+_2$ (**25 µl per each tube)**.

NOTE: It is also necessary to carry out Negative Control of Amplification (NCA) at suspicion on possible contamination.

NCA – Add **25 µl** of **Buffer for elution** to the tube labelled **NCA** (Negative Control of Amplification).

NOTE: Mix the tubes thoroughly by pipetting avoiding foaming

[2] Number of biological samples + 3 controls of RNA extraction (N+3, N – number of biological samples).
[3] Number of biological samples + 3 controls of RNA extraction + 4 DNA calibrators (N+7, N – number of biological samples).
[4] Extraction of 1 stripe with NucliSENS easyMAG automated system (8 tubes).
[5] 12-tube extraction panel.
[6] Extraction of 2 stripes with NucliSENS easyMAG automated system (16 tubes).

8.2.2 Reverse transcription and amplification

1. Create a temperature profile on your instrument as follows.

Table 3

AmpliSens-2 RG program (for rotor-type instruments)

Step	Temperature, °C	Time	Fluorescence detection	Cycles
1	50	15 min	–	1
2	95	15 min	–	1
3	95	5 s	–	5
	60	20 s	–	
	72	15 s	–	
4	95	5 s	–	40
	60	20 s	FAM, JOE, ROX, Cy5	
	72	15 s	–	

Table 4

AmpliSens-2 iQ program (for plate-type instruments)

Step	Temperature, °C	Time	Fluorescence detection	Cycles
1	50	15 min	–	1
2	95	15 min	–	1
3	95	5 s	–	5
	60	20 s	–	
	72	15 s	–	
4	95	5 s	–	40
	60	30 s	FAM, HEX, ROX, Cy5	
	72	15 s	–	

NOTE: The use of either AmpliSens-2 RG or AmpliSens-2 iQ programs allows to simultaneously carry out any test combinations just in one run (for example simultaneously with *HDV* detection tests; *HCV* genotyping and others). In case of one instrument simultaneously performing only the DNA *HBV* detection tests, it is possible to omit the first step of the program (50 °C, 15 min) to spare time.

NOTE: Channels for the ROX and Cy5 fluorophores are activated upon request, when the multiprime-format tests are carried out, which use these channels

3. Adjust the fluorescence channel sensitivity according to the *Important Product Information Bulletin* and Guidelines [2].
4. Insert the tubes into the reaction module of the device.
5. Run the amplification program with fluorescence detection.
6. Analyse results after the amplification program is completed

9. DATA ANALYSIS

Analysis of results is performed by the software of the real-time PCR instrument used by measuring the fluorescence signal accumulation in two channels:

– The signal of the Internal Control cDNA amplification product is detected in the channel for the FAM fluorophore.
– The signal of the *HCV* cDNA amplification product is detected in the channel for the JOE fluorophore.

Results are interpreted by the crossing (or not-crossing) the fluorescence curve with the threshold line set at the specific level (in the middle of the linear fragment of the positive control fluorescence growth in the log scale) that corresponds to the presence (or absence) of the *Ct* values for this sample in the corresponding column of the results grid.

Based on the *Ct* values (the crossing of the fluorescence curve with the threshold line set at the specific level) and on the specified values for the calibrators, PIC1 *HCV* and PIC2 *HCV*, the calibration line is automatically plotted and the values for the number of *HCV* cDNA copies (channel for the JOE fluorophore) and for the number of Internal Control cDNA copies (channel for the FAM fluorophore) in a PCR sample are calculated. The obtained values are used for the *HCV* RNA concentration calculation in test and control samples, using the formula:

$$\frac{\textit{HCV}\text{ cDNA copies per PCR-sample}}{\text{IC cDNA copies per PCR-sample}} \times \text{coefficient A} \times \text{coefficient B} = \text{IU of } \textit{HCV}\text{ RNA/ml of plasma}$$

$$\text{Coefficient A} = \frac{100}{\text{extraction volume, µl}}$$

NOTE: Coefficient A = 1 when calculating PCE-1 and PCE- 2 concentrations

Coefficient B is specified in the *Important Product Information Bulletin* enclosed to the PCR kit and is specific for each lot. It cannot be used with PCR kits of different lots. Coefficient B should be calculated as the result of calibration during the first PCR run (see section 8.1.1 for details).

NOTE: If the result is greater than 100,000,000 IU/ml, then it is interpreted as the **greater than 100,000,000 IU/ml result.** If the obtained value is higher than the linear range, then the sample may be re-tested after 10x dilution; the produced result is multiplied by10.
If the result is less than 300 IU/ml (extraction from 100 µl), or less than 150 IU/ml (extraction from 200 µl), or less than 30 IU/ml (extraction from 1 ml), then it is interpreted as the **less than 300, less than 150, or less than 30 IU/ml result**, respectively.

The result of the analysis is considered reliable only if the results obtained for the Positive and Negative Controls of amplification as well as for the Negative and Positive Controls of extraction are correct (see Table 5)

Table 5

Results for controls

Control	Stage for control	Result of amplification in the channel for fluorophore	
		FAM	JOE
C–	RNA extraction, PCR	Positive (number of IC copies in PCR sample is greater than the boundary value)	Negative
PCE-1	RNA extraction, PCR	Positive (number of IC copies in PCR sample is greater than the boundary value)	Positive (should be within range specified in the Bulletin as a result of calculation with IC copies/ml)
PCE-2	RNA extraction, PCR	Positive (number of IC copies in the PCR sample is greater than the boundary value)	Positive (should be within range specified in the Bulletin as a result of calculation with IC copies/ml)
C+$_1$	PCR	Positive	Positive
C+$_2$	PCR	Positive	Positive
NCA	PCR	Negative	Negative

NOTE: Boundary values and the range of values for **PCE-1 (Positive Control-1-*HCV*)** and **PCE-2 (Positive Control-2-*HCV*)** calculated with IC copies/ml are specified in the *Important Product Information Bulletin* enclosed to the PCR kit.

10. TROUBLESHOOTING

The results are not taken into account in the following cases:

1. The *Ct* value determined for the Positive Controls of Extraction or Amplification (PCE or C+) in the channel for the JOE fluorophore is absent or greater than the specified boundary value. The PCR analysis (beginning with the RNA extraction stage) should be repeated for all the samples in which *HCV* RNA was not detected.
2. The *Ct* value is determined for the Negative Control of Extraction (C–) in the channel for the JOE fluorophore and/or Negative Control of Amplification (NCA) in the channels for the FAM and JOE fluorophores. The PCR analysis (beginning with the RNA extraction stage) should be repeated for all the samples in which *HCV* RNA was detected.
3. The correlation coefficient, R^2 is less than 0.98 when plotting the calibration line, then the amplification and detection for all the samples should be repeated.
4. The calculated concentrations of Positive Control-1 *HCV* and Positive Control-2 *HCV* exceed the range specified in the *Important Product Information Bulletin*. The PCR analysis (beginning with the RNA extraction stage) should be repeated for all the samples.

11. TRANSPORTATION

AmpliSens® *HCV*-Monitor-FRT PCR kit should be transported at 2–8 ºC for no longer than 5 days.

12. STABILITY AND STORAGE

All components of the **PCR kit variant FRT, *HCV*-Q calibration kit** and **Negative Control (C–)** are to be stored at the temperature from minus 24 to minus 16 °C when not in use. All components of the **AmpliSens® *HCV*-Monitor-FRT** PCR kit are stable until the expiry date stated on the label. The shelf life of the reagents before and after the first use is the same, unless otherwise stated.

NOTE: RT-PCR-mix-1-FL *HCV* is to be kept away from light.

NOTE: Do not repeat freeze-thaw cycles more than twice for Positive Control-1-*HCV,* Positive Control-2-*HCV,* PIC1 *HCV,* PIC2 *HCV,* Internal Control *ICZ*-rec. Store the above-mentioned reagents at 2-8 °C for up to 6 months after thawing.

13. SPECIFICATIONS

13.1 Linear measurement range

The linear measurement range of **AmpliSens® *HCV*-Monitor-FRT** PCR kit is specified in the table below.

Table 6

Biological material	Extraction volume, µl	Nucleic acid extraction kit	Linear measurement range, IU/ml
Blood plasma	100	RIBO-prep NucliSENS easyMAG	300 – 100,000,000
	200	MAGNO-sorb	150 – 100,000,000
	1000	MAGNO-sorb NucliSENS easyMAG	30 – 100,000,000

13.2 Analytical specificity

The analytical specificity of **AmpliSens® *HCV*-Monitor-FRT** PCR kit is ensured by the selection of specific primers and probes as well as reaction conditions. The primers and probes were checked for possible homologies to all sequences published in the gene banks by sequence comparison analysis.

The analytical specificity is also ensured by the addition of the genomic DNA/RNA of the following organisms and viruses to the reaction: *hepatitis A virus*; *hepatitis B virus*; *hepatitis D virus*; *human immunodeficiency virus*; *cytomegalovirus*; *Epstein-Barr virus*; *herpes simplex virus* types 1 and 2; *varicella-zoster virus*; *human herpes virus* types 6 and 8; *parvovirus* B19; *tick-borne encephalitis virus*; *West Nile encephalitis virus*; *adenovirus* types 2, 3, and 7; *Escherichia coli*; *Staphylococcus aureus; Streptococcus pyogenes; Streptococcus agalactiae;* and *Homo sapiens*.

No cross-reactions were observed for the abovementioned organisms and viruses.

14. REFERENCES

1. Handbook “Sampling, Transportation, and Storage of Clinical Material for PCR Diagnostics”, developed by Federal State Institute of Science “Central Research Institute of Epidemiology” of Federal Service for Surveillance on Consumers’ Rights Protection and Human Well-Being.
2. Guidelines to **AmpliSens® *HCV*-Monitor-FRT, AmpliSens® *HBV*-Monitor-FRT, and AmpliSens® *HDV*-Monitor-FRT** PCR kits for quantitative detection of *hepatitis C virus* (*HCV)* RNA, *hepatitis B virus* (*HBV)* DNA and *hepatitis D (HDV)* RNA in the biological material by polymerase chain reaction (PCR) with real-time hybridization-fluorescence detection developed by Federal Budget Institute of Science “Central Research Institute for Epidemiology”.

15. QUALITY CONTROL

In compliance with Federal Budget Institute of Science “Central Research Institute for Epidemiology” ISO 13485-Certified Quality Management System, each lot of **AmpliSens® *HCV*-Monitor-FRT** PCR kit has been tested against predetermined specifications to ensure consistent product quality.

List of Changes Made in the Instruction Manual

VER	Location of changes	Essence of changes
05.12.10	Sampling and handling	Sentence «Collect blood samples into tubes with 3% EDTA solution (1 : 20) after overnight fasting» is changed into «Blood samples are taken after overnight fasting into the tube with EDTA solution as anticoagulant».
	Troubleshooting	Items 1 and 2 are corrected in accordance with Russian instruction manual
	Through the text	Corrections through the text
		MAGNO-sorb mention was deleted
13.12.10	Content	DNA calibrators PIC1 *HCV*(C+$_1$) and PIC2 *HCV*(C+$_2$) are changed to DNA calibrators PIC1 *HCV* and PIC2 *HCV*
20.01.11 RT	Data analysis	Item was corrected in accordance with Russian instruction
	Stability and storage	*HCV*-Q calibration kit storage conditions are added
		Phrase about keeping away from light of RT-PCR-mix-1-FL *HCV* is added
		The information about the shelf life of reagents before and after the first use was added
	Cover page	The phrase “For Professional Use Only” was added

VER	Location of changes	Essence of changes
	Intended use	The phrase “The results of PCR analysis are taken into account in complex diagnostics of disease” was added
	Content	New sections “Working Conditions” and “Transportation” were added
		The “Explanation of Symbols” section was renamed to “Key to Symbols Used”
	Key to Symbols Used	The explanation of symbols was corrected
08.07.11 LA	Cover page, text	The name of Institute was changed to Federal Budget Institute of Science “Central Research Institute for Epidemiology”
15.09.11 RT	8. PROTOCOL 8.1. RNA extraction	The information about using RIBO-prep kit was added
02.12.11 VV	3.CONTENT	The mentioning about the PCR kit form that does not correspond to the specified catalogue number is deleted
16.03.12 LA	13.1. Sensitivity	Column “Test material” (blood plasma) is added
		The name of DNA/RNA extraction kit is changed from RIBO-sorb to RIBO-sorb-12
24.06.12 LA	Cover page	Symbol IVD was replaced by RUO symbol
	16. Key to symbols used	
	13.1. Sensitivity	Information related to MAGNO-sorb reagent kit was added to the table of analytical sensitivity
	8.1. RNA extraction	Reference number of MAGNO-sorb reagent kit was added: K2-16-1000-CE
		Information about extraction with MAGNO-sorb is added
29.01.15 PM	Footer	REF R-V1-MC(RG,iQ,Mx,Dt)-CE-B was added
	Content	The form in bulk was added
25.02.15 ME	Through the text	Misprints and inaccuracies was corrected
08.05.15 ME	Text	Clinical material was changed to biological
	1. Intended use	The phrase “The results of PCR analysis are taken into account in complex diagnostics of disease” was changed to “For research use only. Not for diagnostic procedures”
	3. Content	Compact Disk with software was added
	5. General precautions	Corrections in accordance with template
	9. Data analysis	The unit of measurement for coefficient B was deleted. In the table with results for controls criteria for the FAM channel were changed
	13.2. Specificity	The phrase “The clinical specificity of AmpliSens® *HCV*-Monitor-FRT PCR kit was confirmed in laboratory clinical trials” was deleted
02.11.15 ME	8.2. Preparing reverse transcription and PCR	The shelf life of the mixture of RT-PCR-mix-2-FEP/FRT and DTT dried-frozen was added
24.10.17 ME	Cover page	REF R-V1-MC(RG,iQ,Mx,Dt)-CE, REF R-V1-MC(RG,iQ,Mx,Dt)-CE-B” was added
	3. Content	Number of forms was changed from 2 to 3. It is specified that Form 1 is described in the Instruction Manual to AmpliSens® *HCV*-Monitor-FRT PCR kit REF TR-V1-P-M(RG,iQ,Mx,Dt)-CE. The content of form 2 and 3 was specified
28.08.18 EM	3. Content	The colour of reagent was specified
06.05.20 EM	3. Content	Number of forms was changed from 3 to 2.The information about Form 1 REF TR-V1-P-M(RG,iQ,Mx,Dt)-CE was deleted
14.08.20 KK	Through the text	The text formatting was changed
	Footer	The phrase “For research use only. Not for diagnostic procedures” was added
07.10.20 EM	Throughout the text	All the sections were updated according to the template
16.11.20 KK	Footer	The REF R-V1-MC(RG,iQ,Mx,Dt)-CE-B was deleted
	3. Content	The form in bulk was deleted
01.06.21 MM	Footer	The REF R-V1-MC(RG,iQ,Mx,Dt)-CE-B was added
	3. Content	The form in bulk was added

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