

AmpliSens® *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium*-MULTIPRIME-FRT PCR kit



For Professional Use Only

Instruction Manual

KEY TO SYMBOLS USED

	Catalogue number		Contains sufficient for <n> tests
	Batch code		Use-by Date
	Research Use Only		Consult instructions for use
	Version		Keep away from sunlight
	Temperature limit	NCA	Negative control of amplification
	Manufacturer	C-	Negative control of extraction
	Date of manufacture	C+	Positive control of amplification
	Caution	IC	Internal control

1. INTENDED USE

AmpliSens® *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium*-MULTIPRIME-FRT PCR kit is an *in vitro* nucleic acid amplification test for simultaneous detection of *Neisseria gonorrhoeae*, *Chlamydia trachomatis* and *Mycoplasma genitalium* DNA in the biological material (urogenital, rectal, and oropharyngeal swabs; conjunctival discharge; prostate gland secretion; and urine samples) by using real-time hybridization-fluorescence detection of amplified products.

NOTE: For research use only. Not for diagnostic procedures.

2. PRINCIPLE OF PCR DETECTION

Neisseria gonorrhoeae, *Chlamydia trachomatis* and *Mycoplasma genitalium* detection by the multiplex polymerase chain reaction (PCR) is based on the amplification of pathogen genome specific regions using specific primers. In real-time PCR, the amplified product is detected using fluorescent dyes. These dyes are linked to oligonucleotide probes that bind specifically to the amplified product during thermocycling. The real-time monitoring of the fluorescence intensities during the real-time PCR allows the detection of accumulating product without re-opening the reaction tubes after the PCR run.

AmpliSens® *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium*-MULTIPRIME-FRT PCR kit is a qualitative test that contains the Internal Control-FL (Internal Control-FL (IC)). It must be used in the extraction procedure in order to control the extraction process of each individual sample and to identify possible reaction inhibition.

AmpliSens® *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium*-MULTIPRIME-FRT PCR kit uses "hot-start", which greatly reduces the frequency of nonspecifically primed reactions. "Hot-start" is guaranteed by separation of nucleotides and Taq-polymerase by using a wax layer or a chemically modified polymerase (TaqF). Wax melts and reaction components mix only at 95 °C. Chemically modified polymerase (TaqF) is activated by heating at 95 °C for 15 min.

The PCR kit contains the system for prevention of contamination by amplicons using the enzyme uracil-DNA-glycosylase (UDG) and deoxyuridine triphosphate. The enzyme UDG recognizes and catalyzes the destruction of the DNA containing deoxyuridine, but has no effect on DNA containing deoxythymidine. Deoxyuridine is absent in the authentic DNA, but is always present in amplicons, because deoxyuridine triphosphate is a part of dNTP mixture in the reagents for the amplification. Due to the deoxyuridine containing contaminating amplicons are sensitive to the destruction by UDG before the DNA-target amplification. So the amplicons cannot be amplified.

The enzyme UDG is thermolabile. It is inactivated by heating at temperature above 50 °C. Therefore, UDG does not destroy the target amplicons which are accumulated during PCR. The results of amplification are registered in the following fluorescence channels.

Table 1

Channel for fluorophore	FAM	JOE	ROX	Cy5
DNA-target	<i>N.gonorrhoeae</i>	<i>C.trachomatis</i>	<i>M.genitalium</i>	Internal Control-FL
Target gene	16s rRNA gene	cryptic plasmid	gyrB gene	genetically engineered construction

3. CONTENT

AmpliSens® *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium*-MULTIPRIME-FRT PCR kit is produced in 2 forms:

variant FRT-100 F R-B67-F(RG,iQ)-CE;

variant FRT-100 F in bulk¹ R-B67-F(RG,iQ)-CE-B.

Variant FRT-100 F includes:

Reagent	Description	Volume, ml	Quantity
PCR-mix-1-FL <i>N.gonorrhoeae</i> / <i>C.trachomatis</i> / <i>M.genitalium</i>	clear liquid from colorless to light lilac colour	1.2	1 tube
PCR-mix-2-FRT	colorless clear liquid	0.3	2 tubes
Polymerase (TaqF)	colorless clear liquid	0.03	2 tubes
Positive Control complex (C+)	colorless clear liquid	0.2	1 tube
DNA-buffer	colorless clear liquid	0.5	1 tube
Negative Control (C-)*	colorless clear liquid	1.2	1 tube
Internal Control-FL (IC)**	colorless clear liquid	1.0	1 tube

* must be used in the extraction procedure as Negative Control of Extraction.

** add 10 µl of Internal Control-FL (IC) during the DNA extraction procedure directly to the sample/lysis mixture (see DNA-sorb-AM protocol).

Variant FRT-100 F is intended for 110 reactions (including controls).

4. ADDITIONAL REQUIREMENTS

- Transport medium.
- DNA extraction kit.
- Disposable powder-free gloves and laboratory coat.
- Pipettes (adjustable).
- Sterile pipette tips with aerosol filters (up to 100 µl) in tube racks.
- Tube racks.
- Vortex mixer.
- Desktop centrifuge with rotor for 2 ml reaction tubes.
- PCR box.
- Real-time instruments (for example, Rotor-Gene 3000/6000 (Corbett Research, Australia); Rotor-Gene Q (QIAGEN, Germany), iCycler iQ5 (Bio-Rad, USA), Mx3000P (Stratagene, USA)).
- Disposable polypropylene PCR tubes (0.1- or 0.2-ml):
 - a) 0.2-ml PCR tubes with optical transparent domed or flat caps if a plate-type instrument is used;
 - b) 0.2-ml PCR tubes with flat caps or strips of four 0.1-ml Rotor-Gene PCR tubes if a rotor-type instrument is used.
- Refrigerator for 2–8 °C.
- Deep-freezer at the temperature from minus 24 to minus 16 °C.
- Reservoir for used tips.

5. GENERAL PRECAUTIONS

The user should always pay attention to the following:

- Use sterile pipette tips with aerosol barriers and use a new tip for every procedure.
- Store all extracted positive material (specimens, controls and amplicons) away from all other reagents and add it to the reaction mix in a distantly separated facility.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Use disposable protective gloves and laboratory cloths, and protect eyes while samples and reagents handling. Thoroughly wash hands afterwards.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use a kit after its expiration date.
- Dispose of all specimens and unused reagents in accordance with local regulations.
- Samples should be considered potentially infectious and handled in biological cabinet in compliance with appropriate biosafety practices.
- Clean and disinfect all samples or reagents spills using a disinfectant, such as 0.5 % sodium hypochlorite or another suitable disinfectant.
- Avoid samples and reagents contact with the skin, eyes, and mucous membranes. If these solutions come into contact, rinse the injured area immediately with water and seek medical advice immediately.
- Safety Data Sheets (SDS) are available on request.
- Use of this product should be limited to personnel trained in DNA amplification techniques.
- Workflow in the laboratory must be one-directional, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment and reagents in the area where the previous step was performed.



Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

¹ In bulk form contains unlabeled tubes. Tubes with identical reagent are packed in one bag with label.

6. SAMPLING AND HANDLING

Obtaining samples of biological materials for PCR-analysis, transportation and storage is described in manufacturer's handbook [1]. It is recommended to read this handbook before starting work.

AmpliSens® *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium*-MULTIPRIME-FRT PCR kit is intended for analysis of DNA extracted by using DNA extraction kits from the biological material (urogenital swabs; rectal swabs; oropharyngeal swabs; conjunctival discharge and prostate gland secretion; urine samples (use the first part of the stream)).

7. WORKING CONDITIONS

AmpliSens® *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium*-MULTIPRIME-FRT PCR kit should be used at 18–25 °C.

8. PROTOCOL

8.1. DNA extraction

It is recommended to use the following nucleic acid extraction kits:

- DNA-sorb-AM.
- For other nucleic acid extraction kits see Guidelines [2].

The DNA extraction of each test sample is carried out in the presence of **Internal Control-FL (IC)**.

NOTE: Extract DNA according to the manufacturer's protocols.

8.2. Preparing the PCR

8.2.1. Preparing tubes for PCR

The type of tubes depends on the PCR instrument used for analysis. Use disposable filter tips for adding reagents, DNA and control samples into tubes.

The total reaction volume is **25 µl**, the volume of DNA sample is **10 µl**.

1. Thaw the required number of the tubes with **PCR-mix-2-FRT**. Vortex the tubes with **PCR-mix-1-FL *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium***, **PCR-mix-2-FRT**, and **polymerase (TaqF)** and then centrifuge briefly.

Take the required number of tubes/strips for amplification of the DNA obtained from test and control samples.

2. For N reactions (including 2 controls), add to a new tube:

- 10*(N+1) µl of PCR-mix-1-FL *N.gonorrhoeae*/ *C.trachomatis*/ *M.genitalium***,
- 5.0*(N+1) µl of PCR-mix-2-FRT**
- 0.5*(N+1) µl of polymerase (TaqF)**.

Vortex the tube, then centrifuge shortly. Transfer **15 µl** of the prepared mixture to each tube.

Steps 3 and 4 are carried out in both variants.

3. Using tips with aerosol filter, add **10 µl of DNA** obtained at the DNA extraction stage to the prepared tubes.

4. Carry out the control amplification reactions:

NCA – Add **10 µl of DNA-buffer** to the tube labeled NCA (Negative Control of Amplification).

C+ – Add **10 µl of Positive Control complex (C+)** to the tube labeled C+ (Positive Control of Amplification).

C– – Add **10 µl of the sample extracted from the Negative Control (C–) reagent** to the tube labeled C– (Negative control of Extraction).

8.2.2. Amplification

1. Create a temperature profile on your instrument as follows:

Table 2

AmpliSens-1 amplification program						
Step	Rotor-type instruments ²			Plate-type instruments ³		
	Temperature, °C	Time	Cycles	Temperature, °C	Time	Cycles
1	95	15 min	1	95	15 min	1
2	95	5 s	5	95	5 s	5
	60	20 s		60	20 s	
	72	15 s		72	15 s	
3	95	5 s	40	95	5 s	40
	60	20 s Fluorescence acquiring		60	30 s Fluorescence acquiring	
	72	15 s		72	15 s	

Fluorescent signal is detected in the channels for the FAM, JOE, ROX and Cy5 fluorophores.

2. Adjust the fluorescence channel sensitivity according to the *Important Product Information Bulletin* and Guidelines [2].

3. Insert tubes into the reaction module of the device.

4. Run the amplification program with fluorescence detection.

5. Analyze results after the amplification program is completed.

9. DATA ANALYSIS

Analysis of results is performed by the software of the real-time PCR instrument used by measuring fluorescence signal accumulation in four channels:

- The signal of the *Neisseria gonorrhoeae* DNA amplification product is detected in the channel for the FAM fluorophore,
- The signal of the *Chlamydia trachomatis* DNA amplification product is detected in the channel for the JOE fluorophore,
- The signal of the *Mycoplasma genitalium* DNA amplification product is detected in the channel for the ROX fluorophore.
- The signal of the IC DNA amplification product is detected in the channel for the Cy5 fluorophore.

Results are interpreted by the crossing (or not-crossing) the fluorescence curve with the threshold line set at the specific level that corresponds to the presence (or absence) of a *Ct* value of the DNA sample in the corresponding column of the results grid.

Principle of interpretation is the following:

- *Neisseria gonorrhoeae* DNA is **detected** if the *Ct* value is determined in the results grid in the channel for the **FAM** fluorophore. Moreover, the fluorescence curve of the sample should cross the threshold line in the area of typical exponential growth of fluorescence.
- *Chlamydia trachomatis* DNA is **detected** if the *Ct* value is determined in the results grid in the channel for the **JOE** fluorophore. Moreover, the fluorescence curve of the sample should cross the threshold line in the area of typical exponential growth of fluorescence.
- *Mycoplasma genitalium* DNA is **detected** if the *Ct* value is determined in the results grid in the channel for the **ROX** fluorophore. Moreover, the fluorescence curve of the sample should cross the threshold line in the area of typical exponential growth of fluorescence.
- *Neisseria gonorrhoeae*, *Chlamydia trachomatis* and *Mycoplasma genitalium* DNA **are not detected** in a sample if the *Ct* value is not determined (absent) in the channels for the FAM, JOE and ROX fluorophores, whereas the *Ct* value determined in the channel for the Cy5 fluorophore is less than the boundary *Ct* value specified in the *Important Product Information Bulletin*.
- The result is **invalid** if the *Ct* value is not determined (absent) in the channels for the FAM, JOE, ROX and Cy5 fluorophores. In such cases, the PCR analysis should be repeated.

NOTE: Boundary *Ct* values are specified in the *Important Product Information Bulletin* enclosed to the PCR kit. See also Guidelines [2]

The result of the analysis is considered reliable only if the results obtained for both Positive and Negative Controls of Amplification as well as for the Negative Control of Extraction are correct (see Table 3).

Table 3

Results for controls			
Control	Stage for control	Ct value in the channel for fluorophore	
		FAM, JOE, ROX	Cy5
C–	DNA extraction	Absent	< boundary value
NCA	PCR	Absent	Absent
C+	PCR	< boundary value	< boundary value

10. TROUBLESHOOTING

Results of analysis are not taken into account in the following cases:

1. If the *Ct* value determined for the Positive Control of Amplification (C+) in the channels for the FAM, JOE and ROX fluorophores is greater than the boundary *Ct* value or absent, the amplification should be repeated for the samples for which *Ct* value is absent.
2. If the *Ct* value is determined for the Negative Control of Amplification (NCA) and/or Negative Control of Extraction (C–) in the channels for the FAM and/or JOE and/or ROX fluorophores, the PCR analysis starting from the DNA extraction stage should be repeated for all samples for which a *Ct* value was determined in the channels for the FAM and/or JOE and/or ROX fluorophores.

11. TRANSPORTATION

AmpliSens® *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium*-MULTIPRIME-FRT PCR kit should be transported at 2–8 °C for no longer than 5 days.

12. STABILITY AND STORAGE

All components of the **AmpliSens® *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium*-MULTIPRIME-FRT** PCR kit (except for polymerase (TaqF) and PCR-mix-2-FRT) are to be stored at the temperature 2–8 °C when not in use.

All components of the **AmpliSens® *N.gonorrhoeae* / *C.trachomatis* / *M.genitalium*-MULTIPRIME-FRT** PCR kit are stable until the expiry date stated on the label. PCR kit **variant FRT-100 F** can be stored without unpacking at 2 to 8 °C for 3 months from the date of manufacture before opening. Once opened, PCR kit **variant FRT-100 F** should be unpacked in accordance with the storage temperatures for each component. The shelf life of reagents before and after the first use is the same, unless otherwise stated.

NOTE: Polymerase (TaqF) and PCR-mix-2-FRT are to be stored at the temperature from minus 24 to minus 16 °C.

NOTE: PCR-mix-1-FL *N.gonorrhoeae*/ *C.trachomatis*/ *M.genitalium* is to be kept away from light.

² For example, Rotor-Gene 3000, Rotor-Gene 6000, Rotor-Gene Q or equivalent

³ For example, iCycler iQ, iQ5, Mx3000P, Mx3000P or equivalent.

13. SPECIFICATIONS

13.1. Sensitivity

Biological material	Nucleic acid extraction kit	PCR kit	Microorganism	Sensitivity, GE/ml ⁴
Urogenital swabs ⁵	DNA-sorb-AM	variant FRT-100 F	<i>Neisseria gonorrhoeae</i>	5x10 ²
			<i>Chlamydia trachomatis</i>	5x10 ²
			<i>Mycoplasma genitalium</i>	10 ³
Urine ⁶	DNA-sorb-AM	variant FRT-100 F	<i>Neisseria gonorrhoeae</i>	10 ³
			<i>Chlamydia trachomatis</i>	10 ³
			<i>Mycoplasma genitalium</i>	2x10 ³

The analytical sensitivity for each microorganism is preserved in the presence of high DNA concentrations of other analyte microorganisms (up to 10⁹ GE/ml).

13.2. Specificity

The analytical specificity of **AmpliSens® N.gonorrhoeae / C.trachomatis / M.genitalium-MULTIPRIME-FRT** PCR kit is ensured by selection of specific primers and probes as well as stringent reaction conditions. The primers and probes were checked for possible homologies to all sequences published in gene banks by sequence comparison analysis. Nonspecific responses were absent in tests of human DNA samples and DNA samples of the following microorganisms: *Gardnerella vaginalis*, *Lactobacillus* spp., *Escherichia coli*, *Staphylococcus* spp., *Streptococcus* spp., *Candida albicans*, *Ureaplasma urealyticum*, *Ureaplasma parvum*, *Mycoplasma hominis*, *Chlamydia trachomatis*, *Mycoplasma genitalium*, *Neisseria* spp., *Neisseria gonorrhoeae*, *Trichomonas vaginalis*, *Treponema pallidum*, *Toxoplasma gondii*, HSV type 1 and 2, CMV, and HPV.

14. REFERENCES

- Handbook "Sampling, Transportation, and Storage of Clinical Material for PCR Diagnostics", developed by Federal Budget Institute of Science "Central Research Institute for Epidemiology" of Federal Service for Surveillance on Consumers' Rights Protection and Human Well-Being.
- Guidelines "Real-Time PCR Detection of STIs and Other Reproductive Tract Infections", issued by Federal Budget Institute of Science "Central Research Institute for Epidemiology" of Federal Service for Surveillance on Consumers' Rights Protection and Human Well-Being, Moscow.

15. QUALITY CONTROL

In compliance with Federal Budget Institute of Science "Central Research Institute for Epidemiology" ISO 13485-Certified Quality Management System, each lot of **AmpliSens® N.gonorrhoeae / C.trachomatis / M.genitalium-MULTIPRIME-FRT** PCR kit has been tested against predetermined specifications to ensure consistent product quality.

List of Changes Made in the Instruction Manual

VER	Location of changes	Essence of changes
23.06.11 LA	Cover page, text	The name of Institute was changed to Federal Budget Institute of Science "Central Research Institute for Epidemiology"
26.12.12 LA	Cover page	IVD symbol was replaced by RUQ symbol
	16. Key to symbols used	
24.09.15 ME	Through the text	Corrections in accordance with the template
	1. Intended use	The clinical material was specified
	6. Sampling and handling	
	9. Data analysis	The section were rewritten
	10. Troubleshooting	
26.12.17 ME	3. Content	The color of the reagent was specified
03.06.21 EM	2. Principle of PCR detection	The table with targets and the information about the enzyme UDG were added
	Through the text	The text formatting was changed
	Footer	The phrase "Not for use in the Russian Federation" was added
22.06.23 EM	3. Content	REF R-B67-F(RG,iQ)-CE was added
	Footer	

AmpliSens®



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⁴ The quantity of genome equivalents of microorganism per 1 ml of the sample from transport medium.

⁵ Urogenital swabs are to be placed into **Transport Medium for Swabs** or **Transport Medium with Mucolytic Agent**.

⁶ Pretreatment is required.