

AmpliSens® *Helicobacter pylori*-FRT PCR kit



For Professional Use Only

Instruction Manual

KEY TO SYMBOLS USED

	Catalogue number		Contains sufficient for <n> tests
	Batch code		Use-by Date
	Research Use Only		Consult instructions for use
	Version		Keep away from sunlight
	Temperature limit		Keep dry
	Manufacturer	NCA	Negative control of amplification
	Date of manufacture	C-	Negative control of extraction
	Caution	C+	Positive control of amplification
		IC	Internal control

1. INTENDED USE

AmpliSens® *Helicobacter pylori*-FRT PCR kit is an *in vitro* nucleic acid amplification test for qualitative detection of *Helicobacter pylori* DNA in the biological material (tissue (biopsy) material of gastric mucosa, feces, saliva), using real-time hybridization-fluorescence detection of amplified products. The material for PCR is DNA samples extracted from test material.

Indications and contra-indications for use of the reagent kit

The reagent kit is used for the analysis of biological material taken from the persons with suspected helicobacteriosis without distinction of form and presence of manifestation. There are no contra-indications with the exception of cases when the material cannot be taken for medical reasons.

NOTE: For research use only. Not for diagnostic procedures

2. PRINCIPLE OF PCR DETECTION

Principle of testing is based on the DNA extraction from the samples of test material with the exogenous internal control sample (Internal Control-FL (IC)) and simultaneous amplification of DNA fragments of the detected microorganism (*Helicobacter pylori*) and DNA of the internal control with hybridization-fluorescence detection. Exogenous internal control (Internal Control-FL (IC)) allows to control all PCR-analysis stages of each individual sample and to identify possible reaction inhibition.

Amplification of DNA fragments with the use of specific primers and Taq-polymerase enzyme are performed with the DNA samples obtained at the extraction stage. In the real-time PCR, the amplified product is detected with the use of fluorescent dyes. These dyes are linked to oligonucleotide probes, which bind specifically to the amplified product during thermocycling. The real-time monitoring of fluorescence intensities during the real-time PCR allows the detection of accumulating product without re-opening the reaction tubes after the PCR run.

AmpliSens® *Helicobacter pylori*-FRT PCR kit uses "hot-start", which greatly reduces the frequency of nonspecifically primed reactions. "Hot-start" is guaranteed by using chemically modified polymerase (TaqF). The chemically modified polymerase (TaqF) is activated by heating at 95 °C for 15 min.

Variant FRT-50 F contains the system for prevention of contamination by amplicons using the enzyme uracil-DNA-glycosylase (UDG) and deoxyuridine triphosphate (dUTP).

The results of amplification are registered in the following fluorescence channels:

Table 1

Channel for fluorophore	FAM	JOE
DNA-target	IC DNA	<i>Helicobacter pylori</i> DNA
Target gene	Artificially synthesized sequence	16S rDNA

3. CONTENT

AmpliSens® *Helicobacter pylori*-FRT PCR kit is produced in 4 forms:

variant FRT-50 F, H-2831-1-CE

variant FRT-L, H-2832-1-4-CE

variant FRT-50 F in bulk¹, H-2831-1-CE-B

variant FRT-L in bulk¹, H-2832-1-4-CE-B

Variant FRT-50 F includes:

Reagent	Description	Volume, ml	Quantity
PCR-mix-FL <i>Helicobacter pylori</i>	clear liquid from colorless to light lilac colour	0.6	1 tube
PCR-buffer-B	colorless clear liquid	0.3	1 tube
Polymerase (TaqF)	colorless clear liquid	0.03	1 tube
C+ <i>Helicobacter pylori</i>	colorless clear liquid	0.2	1 tube
TE-buffer	colorless clear liquid	0.2	1 tube
Negative Control (C-)*	colorless clear liquid	1.2	1 tube
Internal Control-FL (IC)**	colorless clear liquid	0.5	1 tube
Buffer for elution A***	colorless clear liquid	1.2	3 tubes

* must be used in the extraction procedure as Negative Control of Extraction.

** add 10 µl of Internal Control-FL (IC) during the DNA extraction procedure directly to the sample/lysis mixture (see DNA-sorb-B or RIBO-prep protocol).

*** the reagent is used instead of the reagent for elution of the nucleic acid extraction kit.

Variant FRT-50 F is intended for 55 reactions (including controls).

Variant FRT-L includes:

Reagent	Description	Volume, ml	Quantity
PCR-mix <i>Helicobacter pylori</i> -Lyo	white powder	-	48 tubes of 0.2 ml
C+ <i>Helicobacter pylori</i>	colorless clear liquid	0.2	1 tube
TE-buffer	colorless clear liquid	0.2	1 tube
Negative Control (C-)*	colorless clear liquid	1.2	1 tube
Internal Control-FL (IC)**	colorless clear liquid	0.5	1 tube
Buffer for elution A***	colorless clear liquid	1.2	5 tubes

* must be used in the extraction procedure as Negative Control of Extraction.

** add 10 µl of Internal Control-FL (IC) during the DNA extraction procedure directly to the sample/lysis mixture (see DNA-sorb-B or RIBO-prep protocol).

*** the reagent is used instead of the reagent for elution of the nucleic acid extraction kit.

Variant FRT-L is intended for 48 reactions (including controls).

4. ADDITIONAL REQUIREMENTS

For sampling and pretreatment

- Transport medium for swabs.
- Reagent for pretreatment of viscous fluids (saliva).
- 0.9 % of sodium chloride (sterile saline solution) or phosphate buffered saline (PBS) (137 mM sodium chloride; 2.7 mM potassium chloride; 10 mM sodium monophosphate; 2 mM potassium diphosphate; pH=7,5±0,2).
- Glycerin for long storage of biological material (feces) in conditions of low-temperature freeze.
- Plastic container (50-60 ml) for sampling, storage and transportation of biological samples.
- Disposable screwed or tightly closed polypropylene 1.5-ml and/or 2.0-ml tubes for sampling and pretreatment.
- Teflon-tipped pestle for homogenization in 1.5-ml tubes.
- Sterile pipette tips (up to 1,000 µl) and sterile pipette tips with aerosol filters (up to 100 µl, 200 µl and 1,000 µl).
- Tube racks.
- Vortex mixer.
- PCR box.
- Pipettes (adjustable).
- Refrigerator for 2–8 °C.
- Deep-freezer at the temperature from minus 24 to minus 16 °C.
- Reservoir to throw off and inactivate the material.
- Disposable powder-free gloves and a laboratory coat.

For DNA extraction and amplification

- DNA extraction kit.
- Sterile pipette tips with aerosol filters (up to 10 µl, 100 µl and 200 µl).
- Tube racks.
- Vortex mixer.
- PCR box.
- Real-time instruments (for example, Rotor-Gene Q (QIAGEN, Germany); CFX 96 (Bio-Rad, USA)).
- Disposable polypropylene tubes for variant FRT-50 F:
 - a) screwed or tightly closed 1.5-ml tubes for reaction mixture preparation.
 - b) thin-walled 0.2-ml PCR tubes with optical transparent domed or flat caps or strips of eight 0.2-ml tubes with optical transparent caps if a plate-type instrument is used;
 - c) thin-walled 0.2-ml PCR tubes with flat caps or strips of four 0.1-ml Rotor-Gene PCR tubes if a rotor-type instrument is used.
- Pipettes (adjustable).
- Refrigerator for 2–8 °C.
- Deep-freezer at the temperature from minus 24 to minus 16 °C.
- Reservoir for used tips.
- Disposable powder-free gloves and a laboratory coat.

¹ In bulk form contains unlabeled tubes. Tubes with identical reagent are packed in one bag with label.

5. GENERAL PRECAUTIONS

The user should always pay attention to the following:

- Use sterile pipette tips with aerosol filters and use a new tip for every procedure.
- Store all extracted positive material (specimens, controls and amplicons) away from all other reagents and add it to the reaction mix in a distantly separated facility.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Use disposable protective gloves and laboratory cloths, and protect eyes while samples and reagents handling. Thoroughly wash hands afterwards.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use the PCR kit if the internal packaging was damaged or its appearance was changed.
- Do not use the PCR kit if the transportation and storage conditions according to the Instruction Manual were not observed.
- Do not use a kit after its expiration date.
- Dispose of all specimens and unused reagents in accordance with local regulations.
- Samples should be considered potentially infectious and handled in biological cabinet in compliance with appropriate biosafety practices.
- Clean and disinfect all samples or reagents spills using a disinfectant, such as 0.5 % sodium hypochlorite or another suitable disinfectant.
- Avoid inhalation of vapors, samples and reagents contact with the skin, eyes, and mucous membranes. Harmful if swallowed. If these solutions come into contact, rinse the injured area immediately with water and seek medical advice if necessary.
- While observing the conditions of transportation, operation and storage, there are no risks of explosion and ignition.
- Safety Data Sheets (SDS) are available on request.
- The PCR kit is intended for single use for PCR analysis of specified number of samples (see the section "Content").
- The PCR kit is ready for use in accordance with the Instruction Manual. Use the PCR kit strictly for intended purpose.
- Use of this product should be limited to personnel trained in DNA amplification techniques.
- Workflow in the Laboratory must be one-directional, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment and reagents in the area where the previous step was performed.



Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

6. SAMPLING AND HANDLING

AmpliSens® *Helicobacter pylori*-FRT PCR kit is intended for analysis of the DNA extracted with DNA extraction kits from the biological material (tissue (biopsy) material of gastric mucosa, feces, saliva).

Sampling

Tissue (biopsy) material of gastric mucosa. Tissue material (pieces of tissue no more than 5 mm in diameter) is placed into microtubes with screw caps or into 1.5-2.0-ml tubes with snap caps containing 0.1-0.3 ml of transport medium.

The tissue (biopsy) material of gastric mucosa can be stored before pretreatment:

- at the temperature from 18 to 25 °C – for 6 hours;
- at the temperature from 2 to 8 °C – for 3 days;
- at the temperature from minus 24 to minus 16 °C – for 1 week;
- at the temperature not more than minus 68 °C – for a long time.

Only one freeze-thawing cycle is acceptable.

The pre-frozen material can be transported at 2–8 °C for 1 day.

Feces are taken from a disposable reservoir (for example, a petrie dish, disposable plastic bag) placed into a bed-pan or disposable diapers (for younger children). When using a disposable diaper for children with liquid stool, place a cotton pad into the diaper before the use for obtaining the sufficient quantity of sample.

NOTE: It is forbidden to take feces samples directly from a bed-pan or another reservoir for multiple use (without distinction of disinfection methods).

Using a separate filter tip or disposable spatula transfer about 1.0 g (or 1.0 ml) of the sample into special disposable plastic container.

The feces samples can be stored before pretreatment:

- at the temperature from 18 to 25 °C – for 6 hours;
- at the temperature from 2 to 8 °C – for 3 days;
- at the temperature from minus 24 to minus 16 °C – for 1 week;
- at the temperature not more than minus 68 °C – for a long time.

Only one freeze-thawing cycle is required.

The feces samples can be transported at 2–8 °C for 3 days.

Saliva should be obtained after threefold rinsing the oral cavity with saline solution or boiled water. Saliva is taken into sterile dry disposable 2.0-ml tubes in an amount not less than 0.2-1.0 ml. Close the tube tightly with a cap, avoiding gaps and crumpling of the inner part of the cap. Mark the tube.

The saliva samples can be stored before pretreatment:

- at the temperature from 18 to 25 °C – for 6 hours;
- at the temperature from 2 to 8 °C – for 1 day;
- at the temperature from minus 24 to minus 16 °C – for 1 week;
- at the temperature not more than minus 68 °C – for a long time.

Only one freeze-thaw cycle is allowed.

Pretreatment

Tissue (biopsy) material of gastric mucosa is to be pretreated.

Homogenize a sample of tissue (biopsy) material of gastric mucosa in a tube with transport medium by rubbing it against the walls of the tube using a teflon pestle (new for each sample). A short centrifugation should be performed after the procedure to sediment the drops from the walls of the tube.

The pretreated samples of tissue (biopsy) material of gastric mucosa can be stored before PCR-analysis:

- at the temperature from 18 to 25 °C – for 6 hours;
- at the temperature from 2 to 8 °C – for 1 day;
- at the temperature from minus 24 to minus 16 °C – for 1 week;
- at the temperature not more than minus 68 °C – for a long time.

Feces are to be pretreated.

Fecal suspension preparation:

1. Take the required number of disposable 1.5-ml tubes respectively to the number of samples. Add 1.0 ml of PBS into each tube (use 15-20 % solution of glycerin in PBS when necessary to store the suspension more than 1 day under refrigeration).
2. Using a new one filter tip (or disposable spatula) for each sample add 0.1 g (0.1 ml) of feces into each tube and resuspend thoroughly on vortex due to obtain homogenous suspension. Optimal concentration of suspension is ~ 10 % (by the pellet volume after centrifugation). Sediment the drops from the tube caps by short centrifugation on vortex (no more than 10 sec).

Liquid semitransparent feces are used for express filtration without previous obtaining the suspension.

Express filtration of fecal suspension (for viral and bacterial pathogens detection):

1. For express filtration use two tips up to 1.0 ml (with filter and without filter) and a cut lower part of cotton probe (cotton bud).
2. Put the cut lower part of disposable cotton probe (cotton bud) in the tip without aerosol filter and fix it by pushing into the necked part of the tip.
3. Take 1.0 ml of fecal suspension by the filter tip, put it in the prepared tip with cotton filter and carry out the pressing-filtration into a new disposable tube. In case of difficult filtration it is recommended to decrease the fecal suspension concentration.
4. 100 µl of filtrate is used for DNA extraction.

The pretreated samples of feces suspension can be stored before PCR-analysis:

- at the temperature from minus 24 to minus 16 °C – for 1 week,
- at the temperature not more than minus 68 °C – for a long time.

Only one freeze-thawing cycle is required.

The samples of fecal suspension can be transported at 2–8 °C for 1 day.

Saliva is to be pretreated.

If mucus is present in the sample, Mucolysin reagent should be added to the sample. In order to reduce the viscosity, a threefold amount of Mucolysin reagent respectively to the amount of saliva should be added to the container with saliva and incubated at the room temperature (from 18 to 25 °C), mixing occasionally for 10-20 min (until visual clarification). 100 µl of thin saliva is used for DNA extraction.

The pretreated thin saliva can be stored before PCR-analysis:

- at the temperature from minus 24 to minus 16 °C – for 1 week,
- at the temperature not more than minus 68 °C – for a long time.

Only one freeze-thawing cycle is required.

Interfering substances and limitations of using test material samples

Feces samples taken directly from a bed-pan or another reservoir for multiple use (without distinction of disinfection methods) are inapplicable for analysis.

In order to control the DNA extraction efficiency and possible reaction inhibition the Internal Control (Internal Control-FL (IC)) is used in the PCR kit. The Internal Control is added in each biological sample at the extraction stage. The presence of internal control signal after the amplification testifies the effectiveness of nucleic acid extraction and the absence of PCR inhibitors.

Potential interfering substances

Endogenous and exogenous substances that may be present in the biological material (tissue (biopsy) material of gastric mucosa, feces, saliva) used for the study were selected to assess potential interference.

Model samples of biological material without adding and with the addition of potential interfering substances were tested. The concentration of each potential interfering substance is specified in Table 2. Model samples contained quality control sample (QCS) with *Helicobacter pylori* DNA concentration of 1x10³ GE/ml.

Table 2

Test material	Type of potential interferent	Potential interferent	Tested concentration in a sample	Interference presence
Tissue (biopsy) material of gastric mucosa	Endogenous substances	Whole blood	Up to 40 %	Not detected
		Formalin 10% neutral	Up to 10 %	Not detected
	Exogenous substances	Clarithromycin, 250 mg capsules + Flemoxin Solutab (Amoxicillin 1,000 mg)	Up to 10 mg/ml Clarithromycin + 20 mg/ml Amoxicillin	Not detected
		De-nol (bismuth tripotassium dicitrate 304.6 mg (in terms of bismuth oxide 120 mg))	Up to 12.2 mg/ml bismuth tripotassium dicitrate (or 4.8 mg/ml in terms of bismuth oxide)	Not detected
		Omeprazole-Akrikhin, enterosoluble capsules, omeprazole 20 mg	Up to 1.2 mg/ml	Not detected
Feces	Endogenous substances	Whole blood	Up to 40 %	Not detected
		Fecal fats	Up to 40 %	Not detected
		Mucin (mucus)	Up to 3 %	Detected at concentration greater than 10 mg/ml
	Exogenous substances	'Enterofuril' oral suspension	Up to 4.25 mg/ml	Not detected
		'Enterosgel', oral paste	Up to 174.75 mg/ml	Not detected
		Dextrin (Russia)	Up to 68.6 mg/ml	Not detected
Saliva	Endogenous substances	Whole blood	Up to 10 %	Not detected
		Mucin (mucus)	Up to 1 %	Detected at concentration greater than 1.0 mg/ml
	Exogenous substances	Miramistin 0,01 %	Up to 10 %	Not detected
		Chlorhexidine 0,05 %	Up to 10 %	Not detected

7. WORKING CONDITIONS

AmpliSens® *Helicobacter pylori*-FRT PCR kit should be used at the temperature from 20 to 28 °C and relative humidity from 15 to 75 %.

8. PROTOCOL

8.1. DNA extraction

It is recommended to use the following nucleic acid extraction kits:

- DNA-sorb-B;
- RIBO-prep.

NOTE: Extract the DNA according to the manufacturer's protocol.

The volumes of reagents and samples when the DNA is extracted by DNA-sorb-B and RIBO-prep reagent kits:

The DNA extraction for each test sample is carried out in the presence of **Internal Control-FL (IC)**.

Add 10 µl of **Internal Control-FL (IC)** to each tube.

NOTE: The volume of the test sample is 100 µl.

Add 100 µl of **Negative Control (C-)** into the tube labeled C- (Negative Control of Extraction).

The volume of elution:

- 50 µl (in case of using variant FRT-50 F for the amplification);
- 125 µl (in case of using variant FRT-L for the amplification).

NOTE: Buffer for elution A of **AmpliSens® *Helicobacter pylori*-FRT** PCR kit is used for elution during DNA extraction from the test samples.

8.2. Preparing PCR

8.2.1 Preparing tubes for PCR

The type of tubes depends on the PCR instrument used for analysis. Use disposable filter tips for adding reagents, DNA and control samples into tubes.

Variant FRT-50 F

The total reaction volume is 25 µl, the volume of the DNA sample is 10 µl.

- Calculate the required quantity of each reagent for reaction mixture preparation. For one reaction:
 - 10 µl of PCR-mix-FL *Helicobacter pylori*,
 - 5 µl of PCR-buffer-B,
 - 0.5 µl of Polymerase (TaqF).

Prepare the reaction mixture for the total number of test and control samples plus some extra reaction. See numbers of control samples in item 7. The calculation for the required number of reactions including testing the test and control samples can be performed according to Table 3.

Table 3

Reagent volume per one reaction, µl		Reagent volume for specified number of reactions		
Number of test samples	Number of reactions ²	PCR-mix-FL <i>Helicobacter pylori</i>	PCR-buffer-B	Polymerase (TaqF)
2	6	60	30	3.0
4	8	80	40	4.0
6	10	100	50	5.0
8	12	120	60	6.0
10	14	140	70	7.0
12	16	160	80	8.0
14	18	180	90	9.0
16	20	200	100	10.0
18	22	220	110	11.0
20	24	240	120	12.0
22	26	260	130	13.0
24	28	280	140	14.0
26	30	300	150	15.0
28	32	320	160	16.0

NOTE: Prepare the reaction mixture just before use.

- Thaw the tube with PCR-mix-FL *Helicobacter pylori*. Thoroughly vortex all the reagents of the PCR kit and sediment the drops by vortex.
- In a new tube prepare the reaction mixture. Mix the required quantities of PCR-mix-FL *Helicobacter pylori*, PCR-buffer-B and polymerase (TaqF). Sediment the drops by vortex.
- Take the required number of the tubes or strips for PCR of DNA of test and control samples.
- Transfer 15 µl of the prepared reaction mixture to each tube. Discard the unused reaction mixture.

6. Add 10 µl of DNA samples obtained by extraction of test samples.
NOTE: Avoid transferring the sorbent together with the DNA samples extracted with the reagent kit for extraction on silica gel.
7. Carry out the control amplification reactions:
NCA – Add 10 µl of TE-buffer to the tube labeled NCA (Negative Control of Amplification)
C+ – Add 10 µl of C+ <i>Helicobacter pylori</i> to the tube labeled C+ (Positive Control of Amplification)
C– – Add 10 µl of the sample extracted from the Negative Control (C–) reagent to the tube labeled C– (Negative control of Extraction).

NOTE: Mix the tubes thoroughly by pipetting avoiding foaming.

Variant FRT-L

The total reaction volume is 25 µl, the volume of the DNA sample is 25 µl.

- Take the required number of the tubes with ready-to-use lyophilized reaction mixture PCR-mix *Helicobacter pylori*-Lyo for amplification of DNA from test and control samples (see the number of control samples in point 3).

2. Add 25 µl of DNA samples obtained by extraction of test samples.
NOTE: Avoid transferring the sorbent together with the DNA samples extracted with the reagent kit for extraction on silica gel.
3. Carry out the control amplification reactions:
NCA – Add 25 µl of TE-buffer to the tube labeled NCA (Negative Control of Amplification)
C+ – Add 25 µl of C+ <i>Helicobacter pylori</i> to the tube labeled C+ (Positive Control of Amplification)
C– – Add 25 µl of the sample extracted from the Negative Control (C–) reagent to the tube labeled C– (Negative control of Extraction).

NOTE: Mix the tubes thoroughly by pipetting avoiding foaming.

8.2.2. Amplification

- Create a temperature profile on your instrument as follows³:

Table 4

AmpliSens unified amplification program for rotor-type ⁴ and plate-type ⁵ instruments				
Step	Temperature, °C	Time	Fluorescent signal detection	Cycles
1	50	15 min	–	1
2	95	15 min	–	1
3	95	10 s	–	45
	60	20 s	FAM, JOE	

Any combination of the tests (including tests with reverse transcription and amplification) can be performed in one instrument simultaneously with the use of the unified amplification program. If several tests in "multiplex" format are carried out simultaneously, the detection is enabled in other used channels except for the specified ones. If only the tests for DNA detection are performed in one instrument then the first step of reverse transcription (50 °C – 15 min) can be omitted for time saving.

NOTE:

² Number of reaction including the number of test samples (N), the controls of extraction stage and PCR, and one extra reaction (N+1+2+1).

³ The amplification programs (tables 4, 5) are equivalent for the use with this PCR kit.

⁴ For example, Rotor-Gene Q (QIAGEN, Germany).

⁵ For example, CFX 96 (Bio-Rad, USA).

Table 5

Amplification program for rotor-type³ and plate-type⁴ instruments

Step	Temperature, °C	Time	Fluorescent signal detection	Cycles
1	50	30 min	–	1
2	95	15 min	–	1
3	95	10 s	–	45
	60	25 s	FAM, JOE	
	72	10 s	–	

Fluorescent signal is detected in the channels for the FAM and JOE fluorophores.

The given program (Table 5) can be used for all AmpliSens® PCR kits, intended for detection and differentiation of DNA/RNA of microorganisms causing acute intestinal infections, with the possibility of their combined use in one run of the instrument. If only the tests for DNA detection are performed in one instrument then the first step of reverse transcription (50 °C – 30 min) can be omitted for time saving. If other tests are carried out simultaneously, the detection is also enabled in other used channels.

NOTE:

- Adjust the fluorescence channel sensitivity according to the *Important Product Information Bulletin*.
- Insert tubes into the reaction module of the device.

It is recommended to sediment drops from walls of tubes by short centrifugation before placing them into the instrument.

NOTE:

Insert empty tubes at the edges of reaction module in case of incomplete filling of plate-type instrument.

- Run the amplification program with fluorescence detection.
- Analyze results after the amplification program is completed.

9. DATA ANALYSIS

Analysis of results is performed by the software of the real-time PCR instrument used by measuring fluorescence signal accumulation in three channels:

Table 6

Channel for the fluorophore	FAM	JOE
Amplification product	IC DNA	<i>Helicobacter pylori</i> DNA

Results are interpreted by the crossing (or not-crossing) the S-shaped (sigmoid) fluorescence curve with the threshold line set at the specific level that corresponds to the presence (or absence) of a Ct value of the DNA sample in the corresponding column of the results grid.

Principle of interpretation is the following:

Table 7

Results interpretation		
Ct value in the channel for the fluorophore	JOE	Result
determined or absent	< boundary value	<i>Helicobacter pylori</i> DNA is detected
< boundary value	absent or > boundary value	<i>Helicobacter pylori</i> DNA is NOT detected
absent or > boundary value	absent or > boundary value	Invalid* result

* In case of invalid result, the PCR-analysis should be repeated for the corresponding test sample starting from the DNA extraction stage.

NOTE: Boundary Ct values are specified in the *Important Product Information Bulletin* enclosed to the PCR kit.

The results for controls of extraction and amplification stages must meet the criteria given in Table 8 and in the *Important Product Information Bulletin* enclosed to the PCR kit.

Table 8

Results for controls			
Control	Stage for control	Ct value in the channel for fluorophore	
		FAM	JOE
C–	DNA extraction	< boundary value	absent or > boundary value
NCA	PCR	absent or > boundary value	absent or > boundary value
C+	PCR	< boundary value	< boundary value

Interpretation of some test samples is not possible if the results for the controls deviate from the results specified above (see 10. Troubleshooting).

10. TROUBLESHOOTING

Results of analysis are not taken into account in the following cases:

- The Ct value determined for the Positive Control of amplification (C+) in any of the specified channels for fluorophores (see Table 8) is greater than the boundary value or absent. The results interpretation is not possible for samples in which *Helicobacter pylori* DNA was not detected, the PCR analysis should be repeated for these samples. For the samples in which *Helicobacter pylori* DNA was detected, it is necessary to follow the steps specified in point 4.
- For the Negative Control of Extraction (C–):
 - The Ct value determined in the channel for the JOE fluorophore is less than the boundary value. The contamination of laboratory with amplification products or cross-contamination of reagents / test samples is probable at any stage of PCR analysis. The results interpretation is not possible for the samples in which *Helicobacter pylori* DNA was detected. Measures for detecting and elimination of contamination source must be taken. The PCR analysis (beginning with the DNA extraction stage) should be repeated for these samples;
 - The Ct value determined in the channel for the FAM fluorophore is absent or greater than the boundary value. The results interpretation for the test samples is carried out according to Table 7.
- For the Negative Control of amplification (NCA):
 - The Ct value determined in the channel for the JOE fluorophore is less than the boundary value. The contamination of laboratory with amplification products or cross-contamination of reagents / test samples is probable at any stage of PCR analysis. The results interpretation is not possible for the samples in which *Helicobacter pylori* DNA was detected. Measures for detecting and elimination of contamination source must be taken. The PCR analysis should be repeated for these samples;
 - The Ct value determined in the channel for the FAM fluorophore is less than the boundary value. The contamination of laboratory with amplification products or cross-contamination of reagents / test samples is probable at any stage of PCR analysis. The results interpretation is not possible for the samples in which *Helicobacter pylori* DNA was not detected. Measures for detecting and elimination of contamination source must be taken. The PCR analysis should be repeated for these samples.

4. If the Ct value is determined for the test sample, whereas the area of typical exponential growth of fluorescence (in the "raw" data view mode) is absent (the graphic looks like approximate straight line). It is necessary to check the correctness of selected threshold line level. If the result has been obtained with the correct level of threshold line (base line), the amplification should be repeated for this sample.

11. TRANSPORTATION

AmpliSens® *Helicobacter pylori*-FRT PCR kit should be transported at 2–8 °C for no longer than 5 days.

12. STABILITY AND STORAGE

All components of the **AmpliSens® *Helicobacter pylori*-FRT PCR kit** are to be stored at 2–8 °C when not in use (except for PCR-mix-FL *Helicobacter pylori*, PCR-buffer-B and polymerase (TaqF) included in variant FRT-50 F).

All components of the **AmpliSens® *Helicobacter pylori*-FRT PCR kit** are stable until the expiry date stated on the label. The shelf life of reagents before and after the first use is the same, unless otherwise stated.

NOTE: PCR-mix-FL *Helicobacter pylori*, PCR-buffer-B and polymerase (TaqF) are to be stored at the temperature from minus 24 to minus 16 °C

NOTE: PCR-mix-FL *Helicobacter pylori* is to be kept away from light

NOTE: PCR-mix *Helicobacter pylori*-Lyo is to be kept in packages with a desiccant away from light

13. SPECIFICATIONS

13.1. Analytical sensitivity (limit of detection)

Table 9

Test material	Nucleic acid extraction kit	PCR kit	Analytical sensitivity (limit of detection), GE/ml
Tissue (biopsy) material of gastric mucosa, feces, saliva	DNA-sorb-B, RIBO-prep	variant FRT-50 F, FRT-L	1x10 ³

The claimed features are achieved while respecting the rules specified in the section "Sampling and Handling".

13.2. Analytical specificity

The analytical specificity of **AmpliSens® *Helicobacter pylori*-FRT PCR kit** is ensured by the selection of specific primers and probes as well as stringent reaction conditions. The primers and probes have been checked for possible homologies to all sequences published in gene banks by sequence comparison analysis.

The PCR kit detects *Helicobacter pylori* DNA fragments. The analytical specificity was proved when investigating the DNA of the following microorganisms/strains:

1. Strains from ATCC (American Type Culture Collection, USA) in concentration no less than 1x10⁵ GE/ml:

- *Helicobacter pylori* Strain J99 (ATCC® 700824™) and NCTC 11637 (ATCC® 43504™);
- *Campylobacter jejuni* subsp. *jejuni* (ATCC® 33560™), *Campylobacter coli* (ATCC® 49941™), *Campylobacter fetus* subsp. *fetus* (ATCC® 27374™), *Campylobacter hominis* (ATCC® BAA-381™);
- *Acinetobacter baumannii* (ATCC® 19606™), *Bacteroides fragilis* (ATCC® 25285™), *Bordetella bronchiseptica* (ATCC® 10580™), *Bordetella bronchiseptica* (ATCC® 4617™), *Bordetella pertussis* (ATCC® 9340™), *Candida albicans* (ATCC® 14053™), *Candida guilliermondii* (ATCC® 6260™), *Candida krusei* (ATCC® 14243™), *Clostridium difficile* (ATCC® 9689™), *Clostridium septicum* (ATCC® 12464™), *Corynebacterium jeikeium* (ATCC® 43734™), *Corynebacterium minutissimum* (ATCC® 23348™), *Corynebacterium xerosis* (ATCC® 373™), *Eggerthella lenta* (*Eubacterium lentum*) (ATCC® 43055™), *Enterobacter aerogenes* (ATCC® 13048™), *Enterobacter cloacae* (ATCC® 13047™), *Enterococcus faecalis* (ATCC® 29212™), *Enterococcus faecalis* (*vancomycin resistant*) (ATCC® 51299™), *Enterococcus faecium* (ATCC® 35667™), *Erysipelothrix rhusiopathiae* (ATCC® 19414™), *Escherichia coli* (ATCC® 25922™), *Escherichia coli* (ATCC® 35218™), *Gardnerella vaginalis* (ATCC® 14018™), *Haemophilus influenzae* (ATCC® 33930™), *Haemophilus influenzae* (ATCC® 9006™), *Haemophilus influenzae* (ATCC® 10211™), *Haemophilus parainfluenzae* (ATCC® 7901™), *Klebsiella oxytoca* (ATCC® 49131™), *Klebsiella pneumoniae* subsp. *pneumoniae* (ATCC® 27736™), *Listeria grayi* (*murrayi*) (ATCC® 25401™), *Listeria innocua* (ATCC® 33090™), *Listeria monocytogenes* (ATCC® 7644™), *Moraxella (Branhamella) catarrhalis* (ATCC® 25238™), *Moraxella (Branhamella) catarrhalis* (ATCC® 8176™), *Moraxella catarrhalis* (ATCC® 25240™), *Neisseria meningitidis* (ATCC® 13102™), *Neisseria meningitidis* (ATCC® 13090™), *Neisseria lactamica* (ATCC® 23970™), *Neisseria gonorrhoeae* (ATCC® 19424™), *Neisseria gonorrhoeae* (ATCC® 49926™), *Proteus mirabilis* (ATCC® 12453™), *Proteus vulgaris* (ATCC® 6380™), *Propionibacterium acnes* (ATCC® 11827™), *Pseudomonas aeruginosa* (ATCC® 15442™), *Rhodococcus equi* (ATCC® 6939™), *Salmonella enterica* subsp. *enterica* serovar *Typhimurium* (ATCC® 14028™), *Serratia marcescens* (ATCC® 14756™), *Staphylococcus aureus* subsp. *aureus* (ATCC® 6538P™), *Staphylococcus aureus* subsp. *aureus* (MRSA) (ATCC® 43300™), *Staphylococcus aureus* subsp. *aureus* (ATCC® 29213™), *Staphylococcus aureus* subsp. *aureus* (ATCC® 25923™), *Staphylococcus aureus* subsp. *aureus* (ATCC® 33862™), *Staphylococcus aureus* subsp. *aureus* (MRSA) (ATCC® 33591™), *Staphylococcus aureus* subsp. *aureus* (ATCC® 12600™), *Staphylococcus epidermidis* (ATCC® 12228™), *Staphylococcus haemolyticus* (ATCC® 29970™), *Staphylococcus saprophyticus* (ATCC® 49907™), *Stenotrophomonas maltophilia* (ATCC® 13637™), *Streptococcus agalactiae* (ATCC® 12386™), *Streptococcus agalactiae* (ATCC® 13813™), *Streptococcus dysgalactiae* subsp. *equisimilis* (ATCC® 12388™), *Streptococcus equi* subsp. *equi* (ATCC® 9528™), *Streptococcus bovis* (Group D) (ATCC® 9809™), *Streptococcus mutans* (ATCC® 35668™), *Streptococcus pneumoniae* (ATCC® 49619™), *Streptococcus pneumoniae* (ATCC® 6303™), *Streptococcus pneumoniae* (ATCC® 27336™), *Streptococcus pneumoniae* (ATCC® 6305™), *Streptococcus pyogenes* (ATCC® 19615™), *Streptococcus salivarius* (ATCC® 13419™), *Streptococcus uberis* (ATCC® 700407™), *Vibrio parahaemolyticus* (ATCC® 17802™), *Vibrio vulnificus* (ATCC® 27562™).

2. Human DNA in concentration of 0.2 mg/ml.

The nonspecific reactions were not observed while testing the DNA samples of the above mentioned organisms, as well as human DNA.

The information about interfering substances is specified in the *Interfering substances and limitations of using test material samples*.

13.3. Repeatability and reproducibility

Repeatability and reproducibility were determined by testing positive and negative model samples. Positive samples were a quality control sample (QCS) containing *Helicobacter pylori* DNA with concentration of 1x10³ GE/ml. Negative Control (C-) was used as a negative sample.

Repeatability conditions included testing in the same laboratory, by the same operator, using the same equipment within a short period of time. Reproducibility conditions included testing different lots of PCR kit in different laboratories, by different operators, on different days, using different equipment. The results are presented in Table 10.

Table 10

Sample type	Repeatability		Reproducibility	
	Number of samples	Agreement of results, %	Number of samples	Agreement of results, %
Positive	10	100	40	100
Negative	10	100	40	100

14. REFERENCES

1. Malfcrtheiner P., Megraud F., O'Morain C. et al. European Helicobacter Study Group. Management of Helicobacter pylori infection-the Maastricht V/ Florence Consensus Report// Gut. - 2017. - Vol. 66. - P. 6-30.
2. Sabine Skrebinska, Francis Mégraud, Emilie Bessède. Diagnosis of Helicobacter pylori infection. Helicobacter - 2018 Sep;23 Suppl 1:e12515.
3. Rimbara E., Sasatsu M., Graham D. PCR detection of Helicobacter pylori in clinical samples// Meth Mol Biol. - 2013. - Vol. 943. - P. 279-287.

15. QUALITY CONTROL

In compliance with Federal Budget Institute of Science "Central Research Institute for Epidemiology" ISO 13485-Certified Quality Management System, each lot of the **AmpliSens® *Helicobacter pylori*-FRT PCR kit** has been tested against predetermined specifications to ensure consistent product quality.

List of Changes Made in the Instruction Manual

VER	Location of changes	Essence of changes
14.07.23 EM	3. Content Footer	REF H-2831-1-CE; REF H-2832-1-4-CE were added

AmpliSens®



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