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For Professional Use Only

eSens Acute intestinal infections QL PCR kit

REF ES3541B

Instructions for Use

1 INTENDED USE

eSens Acute intestinal infections QL PCR kit is an *in vitro* nucleic acid amplification test for qualitative detection and differentiation of DNA/RNA of *Shigella* spp., enteroinvasive *E.coli* (*EIEC*), *Salmonella* spp., thermophilic *Campylobacter* spp., group F *Adenoviruses* and group A *Rotaviruses*, *Norovirus* genotype 2, and *Astroviruses* in the clinical material and environmental samples by real-time hybridization-fluorescence detection of amplified products.

NOTE: The results of PCR analysis are taken into account in complex diagnostics of disease.

2 PRINCIPLE OF PCR DETECTION

Detection of acute intestinal infections (AII) by the polymerase chain reaction (PCR) is based on the multiplex amplification of the pathogen genome specific region in two tubes using specific primers. In the real-time PCR, the amplified product is detected with the use of fluorescent dyes. These dyes are linked to oligonucleotide probes, which bind specifically to the amplified product during thermocycling. The real-time monitoring of fluorescence intensities during the real-time PCR allows the detection of accumulating product without re-opening the reaction tubes after the PCR run.

eSens Acute intestinal infections QL PCR kit is a qualitative test that contains the Internal Control (Internal Control -FL (IC)). It must be used in the extraction procedure in order to monitor test stages for each individual sample.

To obtain the complementary DNA (cDNA) on the RNA matrix, a reverse transcription reaction is required.

eSens Acute intestinal infections QL PCR kit uses “hot-start,” which greatly reduces the frequency of nonspecifically primed reactions. “Hot-start” is guaranteed by separation of nucleotides and Taq-polymerase by using a chemically modified polymerase (TaqF). The chemically modified polymerase (TaqF) is activated by heating at 95 °C for 15 min.

The results of amplification are registered in the following fluorescence channels:

Table 1

Channel for fluorophore	**FAM**	**JOE**
Name of PCR-mix	DNA-target	
PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp	*Shigella* spp. / *EIEC* DNA	*Salmonella* spp. DNA
RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus*	*Rotavirus* grA RNA	*Astrovirus* RNA
RT-PCR-mix-1-FEP/FRT *Norovirus* / STI	*Internal Control -FL (IC) cDNA*	*Norovirus* G2 RNA
PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus*	*Campylobacter* spp. DNA	*Adenovirus* grF DNA
Name of PCR-mix	Target gene	
PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp	Ipa H (invasive plasmid antigen)	Ttr (redentase gentiocyanate gene)
RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus*	NSP2	gene for capsid protein
RT-PCR-mix-1-FEP/FRT *Norovirus* / STI	Artificially synthesized sequence	gene for capsid protein
PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus*	23S rRNA	Hexon

3 CONTENT

eSens Acute intestinal infections QL PCR kit (ES3541B) includes:

Reagent	**Description**	**Volume, ml**	**Quantity**
PCR-mix-1-FEP/FRT ***Shigella* spp. / *Salmonella* spp.**	colorless clear liquid	0.6	1 tube
PCR-mix-1-FEP/FRT ***Campylobacter* spp. / *Adenovirus***	colorless clear liquid	0.6	1 tube
RT-PCR-mix-1-FEP/FRT ***Rotavirus / Astrovirus***	clear liquid from colorless to light lilac colour	0.6	1 tube
RT-PCR-mix-1-FEP/FRT ***Norovirus* / STI**	clear liquid from colorless to light lilac colour	0.6	1 tube
PCR-buffer-C	colorless clear liquid	0.3	4 tubes
Polymerase (TaqF)	colorless clear liquid	0.03	4 tubes
TM-Revertase (MMlv)	colorless clear liquid	0.015	4 tubes

RT-G-mix-2	colorless clear liquid	0.015	4 tubes
Positive Control DNA *Shigella sonnei / Salmonella* (C+$_{Shigella\ /\ Salmonella}$)	colorless clear liquid	0.1	1 tube
Positive Control DNA *Campylobacter jejuni / Adenovirus* F-Flu (C+$_{Campylobacter\ /\ Adenovirus}$)	colorless clear liquid	0.1	1 tube
Positive Control cDNA *Rotavirus*-Flu / *Astrovirus* (C+$_{Rotavirus\ /\ Astrovirus}$)	colorless clear liquid	0.1	1 tube
Positive Control cDNA *Norovirus* genotype 2-Flu / STI (C+$_{Norovirus\ genotype\ 2\ /\ STI}$)	colorless clear liquid	0.1	1 tube
TE-buffer	colorless clear liquid	0.5	1 tube
Internal Control -FL (IC)*	colorless clear liquid	0.6	1 tube
Negative Control (C–)**	colorless clear liquid	1.2	1 tube
Buffer for elution B***	colorless clear liquid	1.2	5 tubes

* add **10 µl** of **Internal Control -FL (IC)** during the extraction procedure directly to the sample/lysis mixture.

** must be used in the extraction procedure as Negative control of extraction.

*** must be used in the extraction procedure.

eSens Acute intestinal infections QL PCR kit is intended for 55 reactions (including controls).

4 ADDITIONAL REQUIREMENTS

- RNA/DNA extraction kit.
- Disposable powder-free gloves and a laboratory coat.
- Pipettes (adjustable).
- Sterile RNase-free pipette tips with aerosol filters (up to 200 µl).
- Tube racks.
- Vortex mixer.
- Desktop centrifuge with rotor for 2-ml reaction tubes.
- PCR box.
- Real-time instruments (for example, Rotor-Gene Q (QIAGEN, Germany), CFX 96 Touch, CFX 96 Opus (Bio-Rad, USA), QuantStudio 5 (Thermo Fisher Scientific), or equivalent).
- Disposable polypropylene PCR tubes (0.1- or 0.2-ml) when working with PCR kit variant FRT-100 F:
- 0.2-ml PCR tubes with optical transparent domed or flat caps if a plate-type instrument is used;
- 0.2-ml PCR tubes with flat caps if a rotor-type instrument is used.
- Refrigerator for 2–8 °C.
- Deep-freezer at the temperature from minus 24 to minus 16 °C.
- Reservoir for used tips.

5 GENERAL PRECAUTIONS

The user should always pay attention to the following:

- Use sterile pipette tips with aerosol filters and use a new tip for every procedure.
- Store all extracted positive material (specimens, controls and amplicons) away from all other reagents and add it to the reaction mix in a distantly separated facility.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Use disposable protective gloves and laboratory cloths, and protect eyes while samples and reagents handling. Thoroughly wash hands afterwards.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use a kit after its expiration date.
- Dispose of all specimens and unused reagents in accordance with local regulations.
- Samples should be considered potentially infectious and handled in biological cabinet in compliance with appropriate biosafety practices.
- Clean and disinfect all samples or reagents spills using a disinfectant, such as 0.5 % sodium hypochlorite or another suitable disinfectant.
- Avoid inhalation of vapors, samples and reagents contact with the skin, eyes, and mucous membranes. Harmful if swallowed. If these solutions come into contact, rinse the injured area immediately with water and seek medical advice if necessary.
- Safety Data Sheets (SDS) are available on request.
- Use of this product should be limited to personnel trained in DNA amplification techniques.
- Workflow in the laboratory must be one-directional, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment and reagents in the area where the previous step was performed.

⚠ Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

6 SAMPLING AND HANDLING

NOTE: Obtaining samples of biological materials for PCR-analysis, transportation, and storage are described in the manufacturer's handbook. It is recommended that this handbook is read before starting work.

eSens Acute intestinal infections QL PCR kit is intended for the analysis of RNA/DNA extracted with RNA/DNA extraction kits from the environmental samples (concentrated water samples) and clinical material (faeces samples).

Concentrated water samples are used without treatment.

NOTE: The clinical material must be taken according to state and local authorities' requirements.

7 WORKING CONDITIONS

eSens Acute intestinal infections QL PCR kit should be used at 20–28 °C.

8 PROTOCOL

8.1 RNA/DNA extraction

It is recommended to use the following nucleic acid extraction kits:

- RIBO-prep, **REF** K2-9-Et-50-CE.

The DNA/RNA extraction of each test sample is carried out in the presence of **Internal Control (Internal Control -FL (IC)).**

NOTE: Extract RNA according to the manufacturer's instructions.

NOTE: In case of extracting with the RIBO-prep reagent kit the volume of the **Internal Control (Internal Control -FL (IC))** reagent added to each tube is **10 µl**.

NOTE: Use the Buffer for elution B only from this kit in the procedure of DNA/RNA extraction.

8.2 Preparing reverse transcription and PCR

The total reaction volume is **25 µl**, the volume of cDNA sample is **10 µl**.

8.2.1 Preparing tubes for PCR

Use disposable filter tips for adding reagents, DNA/cDNA and control samples into tubes.

NOTE: Reaction mixture components should be mixed just before analysis with calculating for the required number of reactions (test and control samples) according to Table 1. Note that even for analysis of one test or control DNA/RNA sample, it is necessary to carry out all controls of the RT-PCR stage: Positive Control of Amplification (C+) and Negative Control of Amplification (NCA) for each PCR-mix. It is recommended to mix the reagents for an even reaction number to ensure more exact dosage

NOTE: Carry out all control amplification reactions even while testing only one RNA/DNA sample.

1. Take the required number of tubes including controls. The type of tubes depends on the PCR instrument used for analysis.
2. To prepare the reaction mixture, mix:

- **one of PCR-mix-1 (PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp. or PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus* or RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus* or RT-PCR-mix-1-FEP/FRT *Norovirus* / STI)**
- **PCR-buffer-C**
- **Polymerase (TaqF)**
- **RT-G-mix-2 and TM-Revertase (MMlv) (into the mixture with RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus* or RT-PCR-mix-1-FEP/FRT *Norovirus* / STI) (see Table 2)**

Thoroughly vortex the mixture, make sure that there are no drops on the caps of the tubes.

Table 2

Scheme of reaction mixture preparation

		Reagent volume for specified number of reactions				
Reagent volume for 1 reaction (µl)		**10.00**	**5.00**	**0.25**	**0.50**	**0.25**
The number of test samples	**The number of reactions[1]**	**RT-PCR-mix-1-FEP/FRT**	**RT-PCR-2-FEP/FRT**	**RT-G-mix-2**	**Polymerase (TaqF)**	**TM-Revertase (MMlv)**
2	**6**	**60**	**30**	**1.5**	**3.0**	**1.5**
4	**8**	**80**	**40**	**2.0**	**4.0**	**2.0**

[1] The number of clinical reactions + negative control of extraction + 2 controls of amplification + 1 extra sample (N+1+2+1, N is the number of clinical samples).

6	**10**	**100**	**50**	**2.5**	**5.0**	**2.5**
8	**12**	**120**	**60**	**3.0**	**6.0**	**3.0**
10	**14**	**140**	**70**	**3.5**	**7.0**	**3.5**
12	**16**	**160**	**80**	**4.0**	**8.0**	**4.0**
14	**18**	**180**	**90**	**4.5**	**9.0**	**4.5**
16	**20**	**200**	**100**	**5.0**	**10.0**	**5.0**
18	**22**	**220**	**110**	**5.5**	**11.0**	**5.5**
20	**24**	**240**	**120**	**6.0**	**12.0**	**6.0**
22	**26**	**260**	**130**	**6.5**	**13.0**	**6.5**
24	**28**	**280**	**140**	**7.0**	**14.0**	**7.0**
26	**30**	**300**	**150**	**7.5**	**15.0**	**7.5**
28	**32**	**320**	**160**	**8.0**	**16.0**	**8.0**

3. Transfer **15 µl** of the prepared mixture to the prepared tubes.
4. Add **10 µl** of **RNA/DNA** obtained at the extraction stage to the prepared tubes using tips with aerosol barrier. Dispose of the unused reaction mixture.
5. Carry out the control amplification reactions:

NCA — Add **10 µl of TE-buffer** to the tube labeled **NCA** (Negative Control of Amplification).

C+*Shigella / Salmonella* — Add **10 µl of Positive Control DNA *Shigella sonnei / Salmonella* for PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp.** to the tube labeled **C+*Shigella / Salmonella*** (Positive Control of Amplification).

C+*Campylobacter / Adenovirus* — Add **10 µl of Positive Control DNA *Campylobacter jejuni / Adenovirus* F-Flu for PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus*** to the tube labeled **C+*Campylobacter / Adenovirus*** (Positive Control of Amplification).

C+*Rotavirus / Astrovirus* — Add **10 µl of Positive Control cDNA *Rotavirus*-Flu / *Astrovirus* for RT-PCR-mix-1-FEP/FRT *Rotavirus / Astrovirus*** to the tube labeled **C+*Rotavirus / Astrovirus*** (Positive Control of Amplification).

C+*Norovirus* genotype 2 / STI — Add **10 µl of Positive Control cDNA *Norovirus* genotype 2-Flu/STI for RT-PCR-mix-1-FEP/FRT *Norovirus* / STI** to the tube labeled **C+*Norovirus* genotype 2 / STI** (Positive Control of Amplification).

C– — Add **10 µl of the sample extracted from the Negative Control reagent** to the tube labeled **C–** (Negative control of Extraction).

8.2.2 Amplification

1. Create a temperature profile on your instrument as follows:

Table 3

eSens unified amplification program for rotor-type and plate-type instruments

Step	Temperature, °C	Time	Fluorescent signal detection	Cycles
1	50	15 min	-	1
2	95	15 min	-	1
3	95	10 s	-	45
	60	20 s	FAM, JOE	

NOTE: Any combination of the tests (including tests with reverse transcription and amplification) can be performed in one instrument simultaneously with the use of the unified amplification program. If several tests in "multiprime" format are carried out simultaneously, the detection is enabled in other used channels except for the specified ones.

Amplification program

	Rotor-type Instruments (e.g Rotor-Gene Q or equivalent)			Plate-type Instruments (e.g CFX 96 Touch, CFX 96 Opus, QuantStudio 5 or equivalent)		
Step	Temperature, °C	Time	Cycles	Temperature, °C	Time	Cycles
1	50	30 min	1	50	30 min	1
2	95	15 min	1	95	15 min	1
3	95	10 s	45	95	10 s	45
	60	25 s Fluorescence acquiring		60	25 s Fluorescence acquiring	
	72	10 s		72	10 s	

Fluorescent signal is detected in the channels for FAM and JOE fluorophores.

2. Adjust the fluorescence channel sensitivity.
3. Insert tubes into the reaction module of the device.
4. Run the amplification program with fluorescence detection.
5. Analyze results after the amplification program is completed.

8.3 Instrument Settings

Test settings for rotor-type instruments

Channel	Calibrate/Gain Optimisation	Threshold	Dynamic tube	Slope Correct	More Settings/ Outlier Removal
FAM/Green	from 5FI to 10FI	0.05	On	On	5-10 %
JOE/Yellow	from 5FI to 10FI	0.05	On	On	10 %

Test settings for plate-type instruments

Note: Set the heating/cooling rate ***Ramp Rate* 2,5 °C/s** by clicking the ***Step Options*** button for each step of cycling.

Channel	Threshold
FAM, HEX	Set the threshold line for each channel on the level 10-20 % from the maximum fluorescence level of C+ samples in the last amplification cycle.

9 DATA ANALYSIS

Analysis of results is performed by the software of the real-time PCR instrument used by measuring fluorescence signal accumulation in the channels for FAM and JOE fluorophores:

Results are interpreted by the crossing (or not-crossing) the fluorescence curve with the threshold line set at the specific level that corresponds to the presence (or absence) of a *Ct* value of the DNA sample in the corresponding column of the results grid.

Principle of interpretation is given in Table 4:

The channel for fluorophore	**PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp.**	**PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus***	**RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus***	**RT-PCR-mix-1-FEP/FRT *Norovirus* / STI**
FAM	< boundary value *Shigella* spp. / *EIEC* DNA is detected	< boundary value *Campylobacter* spp. DNA is detected	< boundary value *Rotavirus* grA RNA is detected	< boundary value IC cDNA is detected results are valid
	absent or > boundary value *Shigella* spp. / *EIEC* DNA is not detected*	absent or > boundary value *Campylobacter* spp. DNA is not detected*	absent or > boundary value *Rotavirus* grA RNA is not detected*	absent or > boundary value results are invalid (If the Ct value determined for RT-PCR-mix-1-FEP/FRT Norovirus / STI in the channel for FAM fluorophore is absent or greater than the boundary value, the negative result for other PCR-mixes-1 is invalid. The PCR analysis should be repeated (starting from the DNA/RNA extraction stage) for such test sample)
JOE	< boundary value *Salmonella* spp. DNA is detected	< boundary value *Adenovirus* grF DNA is detected	< boundary value *Astrovirus* grA RNA is detected	< boundary value *Norovirus* G2 RNA is detected
	absent or > boundary value *Salmonella* spp. DNA is not detected*	absent or > boundary value *Adenovirus* grF DNA is not detected*	absent or > boundary value *Astrovirus* grA RNA is not detected*	absent or > boundary value *Norovirus* G2 RNA is not detected*

* If the Ct value determined for RT-PCR-mix-1-FEP/FRT Norovirus / STI in the channel for the FAM fluorophore is less than the boundary value.

NOTE: Boundary *Ct* values are specified in the table below.

The result of the analysis is considered reliable only if the results obtained for Positive and Negative Controls of amplification as well as for the Negative Control of extraction are correct (see Table 5).

Table 5

Results for controls

PCR-mix-1	**Control**	**Stage for control**	***Ct* value in the channel for fluorophore**	
			FAM	**JOE**
All PCR-mixes-1 (except for RT-PCR-mix-1-FEP/FRT *Norovirus* / STI)	C–	RNA/DNA extraction	absent or > boundary value	absent or > boundary value
RT-PCR-mix-1-FEP/FRT *Norovirus* / STI	C–	RNA/DNA extraction	< boundary value	absent or > boundary value
PCR-mix-1 FEP/FRT *Shigella* spp. / *Salmonella* spp.	C+*Shigella / Salmonella*	PCR	< boundary value	< boundary value
PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus*	C+*Campylobacter / Adenovirus*	PCR	< boundary value	< boundary value
RT-PCR-mix-1-FEP/FRT *Rotavirus / Astrovirus*	C+*Rotavirus / Astrovirus*	PCR	< boundary value	< boundary value
RT-PCR-mix-1-FEP/FRT *Norovirus* / STI	C+*Norovirus* genotype 2 / STI	PCR	< boundary value	< boundary value
All PCR-mixes-1	NCA	PCR	absent or > boundary value	absent or > boundary value

Boundary *Ct* values for rotor-type instruments

Clinical samples

PCR-mix-1-FEP/FRT / RT-PCR-mlx-1-FEP/FRT	**Channel for fluorophore**	
	FAM	**JOE**
PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp.	38	38
PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus*	38	38
RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus*	38	38
RT-PCR-mix-1-FEP/FRT *Norovirus* / STI .	38	38

Environmental samples

PCR-mix-1-FEP/FRT / RT-PCR-mix-1-FEP/FRT	**Channel for**	**fluorophore**
	FAM	**JOE**
PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp.	40	40

PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus*	40	40
RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus*	40	40
RT-PCR-mix-1-FEP/FRT *Norovirus* / STI .	38	40

Boundary *Ct* values for plate-type instruments

All types of test samples

PCR-mix-1-FEP/FRT / RT-PCR-mix-1-FEP/FRT	Channel for	
	FAM	JOE
PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp.	40	40
PCR-mix-1-FEPIFRT *Campylobacter* spp. / *Adenovirus*	40	40
RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus*	40	40
RT-PCR-mix-1-FEP/FRT *Norovirus* / STI .	40	40

10 TROUBLESHOOTING

Results of analysis are not taken into account in the following cases:

1. If the *Ct* value determined for the Positive Control of Amplification (C+) in the channels for the FAM or JOE fluorophores is greater than the boundary *Ct* value, the amplification and detection should be repeated for all samples in which the *Ct* value in the channels for FAM and JOE fluorophores was greater than the boundary *Ct* value for required PCR-mix-1.
2. If the *Ct* value determined for the Negative Control of Extraction (C–) (except for PCR-mix-1 FEP/FRT *Norovirus* / STI) and/or Negative Control of Amplification (NCA) in the channels for the FAM or JOE fluorophores is less than the boundary *Ct* value, the PCR analysis (beginning with the DNA/RNA extraction stage) should be repeated for all samples in which DNA/RNA of respective pathogen was detected.

11 TRANSPORTATION

eSens Acute intestinal infections QL PCR kit should be transported at 2–8 °C for no longer than 5 days.

12 STABILITY AND STORAGE

All components of the **eSens Acute intestinal infections QL PCR kit** are to be stored at 2–8 °C when not in use (except for PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp., PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus*, RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus*, RT-PCR-mix-1-FEP/FRT *Norovirus* / STI, PCR-buffer-C, polymerase (TaqF), TM-Revertase (MMlv), and RT-G-mix-2). All components of the **eSens Acute intestinal infections QL PCR kit** are stable until the expiration date on the label. The shelf life of reagents before and after the first use is the same, unless otherwise stated.

NOTE: PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp., PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus*, RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus*, RT-PCR-mix-1-FEP/FRT *Norovirus* / STI, PCR-buffer-C, polymerase (TaqF), TM-Revertase (MMlv), and RT-G-mix-2 are to be stored at the temperature from minus 24 to minus 16 °C.

NOTE: PCR-mix-1-FEP/FRT *Shigella* spp. / *Salmonella* spp., PCR-mix-1-FEP/FRT *Campylobacter* spp. / *Adenovirus*, RT-PCR-mix-1-FEP/FRT *Rotavirus* / *Astrovirus*, and RT-PCR-mix-1-FEP/FRT *Norovirus* / STI are to be kept away from light.

13 SPECIFICATIONS

13.1 Analytical sensitivity

Pathogen	Test material	DNA/RNA extraction kit	PCR kit	Analytical sensitivity, GE/ml (Genome equivalents of the pathogen agent per 1 ml of the sample)
Shigella spp. and enteroinvasive *E. coli* (*EIEC*)	feces	RIBO-prep	eSens Acute intestinal infections QL PCR kit	$1x10^3$
Salmonella spp.	feces	RIBO-prep	eSens Acute intestinal infections QL PCR kit	$1x10^3$
Thermophilic *Campylobacter* spp.	feces	RIBO-prep	eSens Acute intestinal infections QL PCR kit	$1x10^3$
Adenovirus F	feces	RIBO-prep	eSens Acute intestinal infections QL PCR kit	$1x10^4$
Rotavirus A	feces	RIBO-prep	eSens Acute intestinal infections QL PCR kit	$1x10^4$
Norovirus genotype 2	feces	RIBO-prep	eSens Acute intestinal infections QL PCR kit	$5x10^3$
Astrovirus	feces	RIBO-prep	eSens Acute intestinal infections QL PCR kit	$1x10^4$

13.2 Analytical specificity

The analytical specificity of **eSens Acute intestinal infections QL PCR kit** is ensured by selection of specific primers and probes as well as strict reaction conditions. The primers and probes were checked for possible homologies to all sequences deposited in gene banks by sequence comparison analysis.

Specificity was checked in tests of DNA samples of the following microorganisms: *Enterovirus* (*Coxsakie* B1, B2, B3, B4, B5, B6; *Polio (Sabin)* I, II, III); *Adenovirus* strains of serogroups 5 and 7; *Influenza viruses* A (H13N2, H9N2, H8N4, H2N3, H4N6, H11N6, H12N5, H3N8, H1N1, H6N2, H10N7, H5N1), *Influenza virus B*; *Rhinovirus*; *RS virus*; human *Adenoviruses* of types 3, 5, 7, 37, and 40; *Salmonella enteritidis* S-6, *S.choleraesuis* 370, *S.typhimurium* 371, *S.dublin* 373, *S.typhi* C1, *S.abortusovis* 372, and *S.gallinarum-pullorum*; *Shigella flexneri* 851b; *Campylobacter fetus* ssp. *fetus* 25936, and *C.jejuni* ssp. *jejuni* 43435; *Clebsiella* K 65 SW4; *Listeria monocytogenes* USHC 19 and *L.monocytogenes* USHC 52; *Proteus vulgaris* 115/98; *Pseudomonas aeruginosa* DN c1; *Staphylococcus aureus* 653 and *S.aureus* 29112; *Morganella morganii* 619 c 01; *Enterobacter faecalis* 356; as well as 44 *Norovirus* isolates of different gene clusters

of genotypes 1 and 2; 40 *Rotavirus* strains of different [P]G types; 19 *Astrovirus* strains of serogroups 1, 2, 4, 5, and 8; and 15 *Adenovirus* strains of different types and the following bacterial strains (see Table 6).

Table 6

Panel of bacterial pathogens (CDC, USA)

Strain ID	**Organism**	**Strain ID**	**Organism**
K2033	*Salmonella* ser. Grumpensis	K2015	*Salmonella* ser. Oranienburg
K1806	*Salmonella* ser. Newport	AM01144	*Salmonella* ser. Newport
K2077	*Salmonella* ser. Enteriditis	K1810	*Salmonella* ser. Anatum
83-99	*Salmonella* ser. Typhimurium	K1991	*Salmonella* ser. Typhimurium
PS505	*Shigella boydii*	K1898	*Salmonella* ser. Heidelberg
PS408	*Shigella sonnei*	PS555	*Shigella boydii*
B4003	*Shigella sonnei*	F6446	*Shigella dysenteriae*
PS801	*Shigella dysenteriae*	S821X1	*Shigella dysenteriae* type 1
C898	*Shigella dysenteriae* type1	S177X1	*Shigella dysenteriae* type 1
F2035	*Shigella flexneri*	S3314	*Shigella dysenteriae* type 2
E2539-C1	Enterotoxigenic *Escherichia coli* (*ETEC*)	PS071	*Shigella flexneri*
H10407	Enterotoxigenic *Escherichia coli* (*ETEC*)	PS050	*Shigella flexneri*
F1008	Enterotoxigenic *Escherichia coli* (*ETEC*)	F7862	*Shigella flexneri*
EDL 933	Shiga-toxin *E.coli* (*STEC*)	TX1	Enterotoxigenic *Escherichia coli* (*ETEC*)
3543-01	Shiga-toxin *E.coli* (*STEC*)	3525-01	Shiga-toxin *Escherichia coli* (*STEC*)
4752-71	*Proteus vulgaris*	25922	*Escherichia coli* O6:H1
QA/QC	*Citrobacter freundii*	621-64	*Citrobacter freundii*
QA/QC	*Aeromonas*	3910-68	*Aeromonas spp.*
3043-74	*Serratia marcescens*	E9113	*Vibrio cholerae*
QA/QC	*Serratia marcescens*	501-83	*Edwardsiella spp.*
F7894	*Vibrio vulnificus*	587-82	*Providencia stuartii*
F8515	*Yersinia enterocolitica*	27853	*Pseudomonas aeruginosa*
F8510	*Yersinia enterocolitica*	D4989	*Helicobacter cineadi*

Strain ID	Organism	Strain ID	Organism
K4299	*Vibrio parahaemolyticus*	D6827	*Helicobacter pullorum*
F9835	*Vibrio parahaemolyticus*	D5127	*Helicobacter pylori*
K2023	*Salmonella* ser. Kentucky	D2686	*Arcobacter butzleri*
K1684	*Salmonella* O-1, 4, 12 gr. B		

The clinical specificity of **eSens Acute intestinal infections QL PCR kit** was confirmed in laboratory clinical trials.

14 QUALITY CONTROL

The production process, including batch release, is carried out in accordance with an established quality management system certified according to ISO 13485.

KEY TO SYMBOLS USED

REF	Catalogue number		Contains sufficient for <n> tests
LOT	Batch code		Use-by Date
IVD	In vitro diagnostic medical device		Consult instructions for use
VER	Version		Keep away from sunlight
	Temperature limit	NCA	Negative control of amplification
	Manufacturer	C–	Negative control of extraction
	Date of manufacture	C+*Shigella / Salmonella*, C+*Campylobacter / Adenovirus*, C+*Rotavirus / Astrovirus*, C+*Norovirus* genotype 2 / STI	Positive control of amplification
	Caution	IC	Internal control

List of Changes Made in the Instruction Manual

VER	Location of changes	Essence of changes
01_04/2022		
02_04/2025	Through the text	The name and volume of the component were modified.
	7 WORKING CONDITIONS	This section was modified.

	8.2 Preparing reverse transcription and PCR	This section was modified.

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