



For Professional Use Only

# eSens *Candida albicans* QL PCR kit

**REF ES3006A**

## Instructions for Use

### 1 INTENDED USE

**eSens *Candida albicans* QL PCR kit** is an *in vitro* nucleic acid amplification test for qualitative detection of *Candida albicans* DNA in the clinical material (urogenital and pharyngeal swabs; urine samples) using real-time hybridization-fluorescence detection of amplified products.

NOTE: The results of PCR analysis are taken into account in complex diagnostics of disease.

### 2 PRINCIPLE OF PCR DETECTION

*Candida albicans* detection by the polymerase chain reaction (PCR) is based on the amplification of a pathogen genome specific region using specific *Candida albicans* primers. In real-time PCR, the amplified product is detected with the use of fluorescent dyes. These dyes are linked to oligonucleotide probes, which bind specifically to the amplified product during thermocycling. The real-time monitoring of fluorescence intensities during the real-time PCR allows the detection of accumulating product without re-opening the reaction tubes after the PCR run.

**eSens *Candida albicans* QL PCR kit** is a qualitative test that contains the Internal Control (**Internal Control-FL (IC)**). It must be used in the extraction procedure in order to control the extraction process of each individual sample and to identify possible reaction inhibition.

**eSens *Candida albicans* QL PCR kit** uses “hot-start,” which greatly reduces the frequency of nonspecifically primed reactions. “Hot-start” is guaranteed by using chemically modified polymerase (TaqF). The chemically modified polymerase (TaqF) is activated by heating at 95 °C for 15 min.

**eSens *Candida albicans* QL PCR kit** contains the system for prevention of contamination by amplicons using the enzyme uracil-DNA-glycosylase (UDG) and dUTP. The enzyme UDG recognizes and catalyzes the destruction of the DNA containing deoxyuridine, but has no effect on DNA containing deoxythymidine. Deoxyuridine is absent in the authentic DNA, but is always present in amplicons, because dUTP is a part of dNTP mixture in the reagents for the amplification. Due to the deoxyuridine containing contaminating amplicons are sensitive to the destruction by UDG before the DNA-target amplification. So the amplicons cannot be amplified.

The enzyme UDG is thermolabile. It is inactivated by heating at temperature above 50 °C. Therefore, UDG does not destroy the target amplicons which are accumulated during PCR.

The results of amplification are registered in the following fluorescence channels:

**Table 1**

Channel for fluorophore	FAM	JOE
DNA-target	<i>Candida albicans</i> DNA	Internal Control-FL (IC) DNA
Target gene	ITS-2 gene	Artificially synthesized sequence

### 3 CONTENT

**eSens Candida albicans QL PCR kit** contains:

Reagent	Description	Volume, ml	Quantity
<b>PCR-mix-1-FL <i>Candida albicans</i></b>	clear liquid from colorless to light lilac colour	1.2	1 tube
<b>PCR-mix-2-FRT</b>	colorless clear liquid	0.3	2 tubes
<b>Polymerase (TaqF)</b>	colorless clear liquid	0.03	2 tubes
<b>Positive Control complex (C+)</b>	colorless clear liquid	0.2	1 tube
<b>DNA-buffer</b>	colorless clear liquid	0.5	1 tube
<b>Negative Control (C-)*</b>	colorless clear liquid	1.2	1 tube
<b>Internal Control-FL (IC)**</b>	colorless clear liquid	1.0	1 tube

\* must be used in the extraction procedure as Negative Control of Extraction.

\*\* add **10 µl** of **Internal Control-FL (IC)** during the DNA extraction procedure directly to the sample/lysis mixture.

**eSens Candida albicans QL PCR kit** is intended for 110 reactions (including controls).

### 4 ADDITIONAL REQUIREMENTS

- Transport medium.
- DNA extraction kit.
- Disposable powder-free gloves and a laboratory coat.
- Pipettes (adjustable).
- Sterile pipette tips with aerosol filters (up to 200 µl).
- Tube racks.
- Vortex mixer.
- Desktop centrifuge with rotor for 2-ml reaction tubes.
- PCR box.
- Real-time instruments (for example, Rotor-Gene Q (QIAGEN, Germany), CFX 96 Touch, CFX 96 Opus (Bio-Rad, USA), QuantStudio 5 (Thermo Fisher Scientific), or equivalent).
- Disposable polypropylene tubes:
  - thin-walled 0.2-ml PCR tubes with domed caps if a plate-type instrument is used;

- thin-walled 0.2-ml PCR tubes with flat caps or strips of four 0.1-ml Rotor-Gene PCR tubes if a rotor-type instrument is used.
- Refrigerator for 2–8 °C.
- Deep-freezer at the temperature from minus 24 to minus 16 °C.
- Reservoir for used tips.

## 5 GENERAL PRECAUTIONS

The user should always pay attention to the following:

- Use sterile pipette tips with aerosol filters and use a new tip for every procedure.
- Store all extracted positive material (specimens, controls and amplicons) away from all other reagents and add it to the reaction mix in a distantly separated facility.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Use disposable protective gloves and laboratory cloths, and protect eyes while samples and reagents handling. Thoroughly wash hands afterwards.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use a kit after its expiration date.
- Dispose of all specimens and unused reagents in accordance with local regulations.
- Samples should be considered potentially infectious and handled in biological cabinet in compliance with appropriate biosafety practices.
- Clean and disinfect all samples or reagents spills using a disinfectant, such as 0.5 % sodium hypochlorite or another suitable disinfectant.
- Avoid inhalation of vapors, samples and reagents contact with the skin, eyes, and mucous membranes. Harmful if swallowed. If these solutions come into contact, rinse the injured area immediately with water and seek medical advice if necessary.
- Safety Data Sheets (SDS) are available on request.
- Use of this product should be limited to personnel trained in DNA amplification techniques.
- Workflow in the laboratory must be one-directional, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment and reagents in the area where the previous step was performed.



Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

## 6 SAMPLING AND HANDLING

**eSens Candida albicans QL PCR kit** is intended for analysis of the DNA extracted with DNA extraction kits from the clinical material (urogenital and pharyngeal swabs; urine samples (sediment of the first portion of the morning specimen)).

## 7 WORKING CONDITIONS

**eSens Candida albicans QL PCR kit** should be used at 18–25 °C.

## 8 PROTOCOL

### 8.1 DNA extraction

Any commercial nucleic acid extraction kit, if IVD-CE validated for the indicated specimen types, could be used.

#### Ecoli Dx, s.r.o. recommends:

- For the manual extraction

- **DNA-sorb-AM** (K1-12-100-CE)

- For the automatic extraction

- **ePure STD DNA Extraction Kit (E2007)**

The DNA extraction of each test sample is carried out in the presence of **Internal Control-FL (IC)**.

Please carry out nucleic acid extraction according to the manufacture´s instruction.

### 8.2 Preparing PCR

#### 8.2.1 Preparing tubes for PCR

The type of tubes depends on the PCR instrument used for analysis. Use disposable filter tips for adding reagents, DNA and control samples into tubes.

The total reaction volume is **25 µl**, the volume of DNA sample is **10 µl**.

1. Thaw the tube with **PCR-mix-2-FRT**. Vortex the tubes with **PCR-mix-1-FL *Candida albicans*, PCR-mix-2-FRT** and **Polymerase (TaqF)** and sediment the drops by short centrifugation (1-2 s). Take the required number of tubes/strips for amplification of DNA from clinical and control samples.
2. For N reactions (including 2 controls of amplification), add to a new tube:  
**10·(N+1) µl** of **PCR-mix-1-FL *Candida albicans*,**  
**5.0·(N+1) µl** of **PCR-mix-2-FRT,**  
**0.5·(N+1) µl** of **polymerase (TaqF).**  
Mix the prepared mixture and sediment the drops by short centrifugation (1-2 s). Transfer **15 µl** of the prepared mixture into each tube.
3. Add **10 µl** of **DNA samples** obtained at the DNA extraction stage.
4. Carry out the control amplification reactions:

<b>NCA</b>	Add 10 µl of DNA-buffer to the tube labeled NCA (Negative Control of Amplification).
<b>C+</b>	Add 10 µl of Positive Control complex (C+) to the tube labeled C+ (Positive Control of Amplification).
<b>C-</b>	Add 10 µl of sample extracted from the Negative Control (C-) reagent to the tube labeled C- (Negative Control of Extraction).

#### 8.2.2 Amplification

1. Create a temperature profile on your instrument as follows:

Table 2

## eSens-1 amplification program

	Rotor-type Instruments (e.g Rotor-Gene Q or equivalent)			Plate-type Instruments (e.g CFX 96 Touch, CFX 96 Opus, QuantStudio 5 or equivalent)		
Step	Temperature, °C	Time	Cycles	Temperature, °C	Time	Cycles
1	95	15 min	1	95	15 min	1
2	95	5 s	5	95	5 s	5
	60	20 s		60	20 s	
	72	15 s		72	15 s	
3	95	5 s	40	95	5 s	40
	60	20 s		60	30 s	
		Fluorescence acquiring			Fluorescence acquiring	
72	15 s	72	15 s			

Fluorescent signal is detected in the channels for the FAM and JOE fluorophores. Other channels are enabled if several tests are simultaneously carried out in a single run.

2. Adjust the fluorescence channel sensitivity.
3. Insert tubes into the reaction module of the device.
4. Run the amplification program with fluorescence detection.

Analyze results after the amplification program is completed.

### 8.3 Instrument Settings

#### Test settings for rotor-type instruments

Channel	Calibrate/Gain Optimisation	Threshold	Dynamic tube	Slope Correct	More Settings/ Outlier Removal
FAM/Green	from 5 FI to 10 FI	0.1	On	Off	0%
JOE/Yellow	from 4 FI to 8 FI	0.1	On	Off	5%

#### Test settings for plate-type instruments

Set the heating/cooling **Ramp Rate 2,5 °C/s**.

Channel	Threshold
FAM JOE/HEX	For each channel in <b>Log Scale</b> set the threshold line at the level of 10-20 % of maximum fluorescence obtained for the Positive Control of Amplification (C+) in the last amplification cycle

## 9 DATA ANALYSIS

Analysis of results is performed by the software of the real-time PCR instrument used by measuring fluorescence signal accumulation in two channels:

- The signal of the *Candida albicans* DNA amplification product is detected in the channel for the FAM fluorophore.
- The signal of the IC DNA amplification product is detected in the channel for the JOE fluorophore.

Results are interpreted by the crossing (or not-crossing) the fluorescence curve with the threshold line set at the specific level that corresponds to the presence (or absence) of a Ct value of the DNA sample in the corresponding column of the results grid.

Principle of interpretation is the following:

- *Candida albicans* DNA is **detected** in a sample if the Ct value is determined in the results grid in the channel for the FAM fluorophore. Moreover, the fluorescence curve of the sample should cross the threshold line in the area of typical exponential growth of fluorescence.
- *Candida albicans* DNA is **not detected** if the Ct value is not determined (absent) in the results grid (the fluorescence curve does not cross the threshold line) in the channel for the FAM fluorophore, whereas the Ct value determined in the results grid in the channel for the JOE fluorophore does not exceed the specified boundary value.
- The result of analysis is **invalid** if the Ct value is not determined in the results grid (absent) in the channel for the FAM fluorophore, whereas the Ct value in the channel for the JOE fluorophore is not determined (absent) or exceeds the specified boundary value. In such cases PCR should be repeated for this sample.

**The result of the analysis is considered reliable only if the results obtained for the Positive and Negative Controls of amplification as well as for the Negative Control of extraction are correct (see Table 3 and 4).**

**Table 3**

**Results for controls**

Control	Stage for control	Ct value in the channel for fluorophore	
		FAM	JOE
C-	DNA extraction	Absent	<boundary value
NCA	PCR	Absent	Absent
C+	PCR	<boundary value	<boundary value

Table 4

Boundary Ct values

Sample	Rotor-type instrument		Plate-type instrument	
	Channel for fluorophore			
	FAM	JOE	FAM	JOE
C+	30	30	33	33
NGA	Ct is absent		Ct is absent	
C-	Ct is absent	30	Ct is absent	33
Test samples	-	30	-	33

## 10 TROUBLESHOOTING

Results of analysis are not taken into account in the following cases:

- The Ct value determined for the Positive Control of amplification (C+) in the channel for the FAM fluorophore is greater than the specified boundary value or absent. The amplification should be repeated for all the samples in which the *Candida albicans* DNA was **not detected**.
- The Ct value is determined for the Negative Control of Extraction (C-) and/or the Negative Control of Amplification (NCA) in the channel for the FAM fluorophore. The PCR analysis (beginning with the DNA extraction stage) should be repeated for all samples in which *Candida albicans* DNA **was detected**.

## 11 TRANSPORTATION

**eSens Candida albicans QL PCR kit** should be transported at 2–8 °C for no longer than 5 days.

## 12 STABILITY AND STORAGE

All components of the **eSens Candida albicans QL PCR kit** are to be stored at 2–8 °C when not in use (except for polymerase (TaqF) and PCR-mix-2-FRT). All components of the **eSens Candida albicans QL PCR kit** are stable until the expiration date on the label. PCR kit can be stored without unpacking at 2 to 8 °C for 3 months from the date of manufacture before opening. Once opened, PCR kit should be unpacked in accordance with the storage temperatures for each component. The shelf life of opened reagents is the same as that of unopened reagents, unless otherwise stated.

NOTE: Polymerase (TaqF) and PCR-mix-2-FRT are to be stored at temperature from minus 24 to minus 16 °C.

NOTE: PCR-mix-1-FL *Candida albicans* should be kept away from light.

## 13 SPECIFICATIONS

### 13.1 Sensitivity

The analytical sensitivity of **eSens Candida albicans QL PCR kit** is following:

Clinical material	Transport medium	Nucleic acid extraction kit	Sensitivity, GE/ml*
Urogenital swabs	Transport Medium for Swabs or Transport Medium with Mucolytic Agent	DNA-sorb-AM ePure STD DNA Extraction kit	1x10 <sup>3</sup>
Urine (pretreatment is required)	–	DNA-sorb-AM ePure STD DNA Extraction kit	2x10 <sup>3</sup>

\*The quantity of genome equivalents of microorganism per 1 ml of the sample from transport medium.

### 13.2 Specificity

The analytical specificity of **eSens Candida albicans QL PCR kit** is ensured by selection of specific primers and probes as well as stringent reaction conditions. The primers and probes were checked for possible homologies to all sequences published in gene banks by sequence comparison analysis.

Nonspecific reactions were absent while testing human DNA samples and DNA panel of the following microorganisms: *Gardnerella vaginalis*, *Lactobacillus* spp., *Escherichia coli*, *Candida glabrata*, *Candida krusei*, *Staphylococcus aureus*, *Streptococcus pyogenes*, *Streptococcus agalactiae*, *Mycoplasma hominis*, *Ureaplasma urealyticum*, *Ureaplasma parvum*, *Mycoplasma genitalium*, *Neisseria flava*, *Neisseria subflava*, *Neisseria sicca*, *Neisseria mucosa*, *Neisseria gonorrhoeae*, *Trichomonas vaginalis*, *Treponema pallidum*, *Toxoplasma gondii*, HSV of 1 and 2 types, CMV and HPV.

The clinical specificity of **eSens Candida albicans QL PCR kit** was confirmed in laboratory clinical trials.

## 14 QUALITY CONTROL

The production process, including batch release, is carried out in accordance with an established quality management system certified according to ISO 13485.

## 15 KEY TO SYMBOLS USED

 REF	Catalogue number		Caution
 LOT	Batch code		Contains sufficient for <n> tests
 IVD	<i>In vitro</i> diagnostic medical device		Use-by Date
 VER	Version		Consult instructions for use
	Temperature limit		Keep away from sunlight
	Manufacturer	NCA	Negative control of amplification
	Date of manufacture	C-	Negative control of extraction
 EC REP	Authorized representative in the European Community	C+	Positive control of amplification
		IC	Internal control

### List of Changes Made in the Instruction Manual

VER	Location of changes	Essence of changes
01_04/2022		

Ecoli Dx, s.r.o. , Purkyňova 74/2



110 00 Praha 1, Česká republika  
Tel: +420 325 209 912

Mobil: +420 739 802 523